

Response to short-term deprivation of the human adult visual cortex measured with 7T BOLD

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1 Abstract

2 Sensory deprivation during the post-natal “critical period” leads to structural reorganization of the
3 developing visual cortex. In adulthood, the visual cortex retains some flexibility and adapts to
4 sensory deprivation. Here we show that short-term (2h) monocular deprivation in adult humans
5 boosts the BOLD response to the deprived eye, changing ocular dominance of V1 vertices,
6 consistent with homeostatic plasticity. The boost is strongest in V1, present in V2, V3 & V4 but
7 absent in V3a and hMT+. Assessment of spatial frequency tuning in V1 by a population Receptive-
8 Field technique shows that deprivation primarily boosts high spatial frequencies, consistent with a
9 primary involvement of the parvocellular pathway. Crucially, the V1 deprivation effect correlates
10 across participants with the perceptual increase of the deprived eye dominance assessed with
11 binocular rivalry, suggesting a common origin. Our results demonstrate that visual cortex,
12 particularly the ventral pathway, retains a high potential for homeostatic plasticity in the human
13 adult.

14 Introduction

15 To interact efficiently with the world, our brain needs to fine-tune its structure and function,
16 adapting to a continuously changing external environment. This key property of the brain, called

17 *neuroplasticity*, is most pronounced early in life, within the so called *critical period*, when
18 abnormal experience can produce structural changes at the level of the primary sensory cortex
19 (Berardi, Pizzorusso, & Maffei, 2000; Hubel & Wiesel, 1970; Hubel, Wiesel, & LeVay, 1977;
20 Wiesel & Hubel, 1963). During development, occluding one eye for a few days induces a dramatic
21 and permanent reorganization of ocular dominance columns (the V1 territory representing each eye)
22 in favor of the open eye (Berardi, Pizzorusso, & Maffei, 2000; Gordon & Stryker, 1996; Hubel &
23 Wiesel, 1970; Hubel, Wiesel, & LeVay, 1977; Wiesel & Hubel, 1963), while the deprived eye
24 becomes functionally blind or very weak. These forms of structural plasticity have been
25 documented in animal models, including non-human primates (Gordon & Stryker, 1996; Kiorpes et
26 al., 1998; Levi & Carkeet, 1993; Wiesel & Hubel, 1963). A corresponding perceptual phenomenon
27 known as amblyopia is observed in humans, and may result from exposing infants to monocular
28 deprivation during the critical period, e.g. due to cataracts (Braddick & Atkinson, 2011; Maurer,
29 Mondloch, & Lewis, 2007). In infants, even a partial deprivation produced by optical defects like
30 astigmatism and myopia leads to a permanent acuity loss that cannot be compensated in adulthood,
31 even after correction the optical aberrations (Freeman & Thibos, 1975) through Adaptive Optics
32 (Rossi et al., 2007). Hebbian plasticity, endorsed by Long-Term synaptic Potentiation and
33 Depression (LTP/LTD) of early stage of cortical processing, underlies these changes in animal
34 models and probably also in humans.

35 After the closure of the critical period, structural changes of V1 resulting from Hebbian plasticity
36 are not typically observed (Mitchell & Sengpiel, 2009; Sato & Stryker, 2008). However, there is
37 evidence that Hebbian plasticity can be restored in adult animal models under special conditions,
38 associated with manipulation of the excitability of the visual cortex (Fong et al., 2016; Fregnac et
39 al., 1988; He et al., 2006; Maya Vetencourt et al., 2008).

40 Besides Hebbian plasticity, other mechanisms can reshape primary visual cortex processing both
41 within and outside the critical period. At the cellular level, there is evidence for homeostatic
42 plasticity, which increases the gain of cortical responses following sensory deprivation; for
43 example, after a brief monocular deprivation, the response gain of the deprived eye increases
44 (Maffei, Nelson, & Turrigiano, 2004). This is interpreted as an homeostatic response to preserve
45 cortical excitability in spite of the synaptic depression produced by Hebbian plasticity, suggesting a
46 close link between these two types of plasticity (Maffei & Turrigiano, 2008; Turrigiano, 2012)
47 (Mrsic-Flogel et al., 2007; Turrigiano & Nelson, 2004).

48 In adult animal models and humans, there is clear evidence for both functional plasticity and for
49 stability of the early sensory cortex (Baseler et al., 2002; Baseler et al., 2011; Wandell & Smirnakis,
50 2009). Functional changes have been observed with perceptual learning (Doshier & Lu, 2017; Fahle
51 & Poggio, 2002; Fiorentini & Berardi, 1980; Karni & Sagi, 1991; Karni & Sagi, 1993; Watanabe &
52 Sasaki, 2015), adaptation that, in some cases, may be very long-lasting, (McCollough, 1965), and
53 short-term visual deprivation (Binda & Lunghi, 2017; Kwon et al., 2009; Lunghi, Berchicci, et al.,
54 2015; Lunghi, Burr, & Morrone, 2011; Lunghi, Burr, & Morrone, 2013; Mon-Williams et al., 1998;
55 Zhang et al., 2009 ; Zhou, Clavagnier, & Hess, 2013; Zhou, Reynaud, & Hess, 2014). The effect of
56 short-term deprivation in adults is paradoxical, boosting the perception of the deprived stimulus –
57 opposite to the long-term deprivation effects during development. One of the first examples of
58 short-term deprivation in adults is by Mon-Williams et al. (1998), who found that thirty minutes of
59 simulated myopia (optical blur achieved by wearing a +1D lens) was followed by a transient
60 improvement of visual acuity – opposite to the long-lasting acuity deficit produced by early onset
61 myopia (Rossi et al., 2007). Contrast attenuation for 4 hours leads to improved contrast
62 discrimination thresholds and enhanced BOLD response in V1/V2 (Kwon et al., 2009). A few hours
63 deprivation of one cardinal orientation leads to enhanced sensitivity to the deprived orientation
64 (Zhang et al., 2009) – opposite to the reduced sensitivity to orientations deprived during
65 development, e.g. due to astigmatism. Similarly, two hours of monocular contrast deprivation is
66 followed by a transient boost of the deprived eye (Binda & Lunghi, 2017; Lunghi, Berchicci, et al.,
67 2015; Lunghi, Burr, & Morrone, 2011; Lunghi, Burr, & Morrone, 2013; Lunghi, Emir, et al., 2015;
68 Zhou, Clavagnier, & Hess, 2013; Zhou, Reynaud, & Hess, 2014) and an enlargement of the
69 deprived-eye representation at the level of V1 in non-human primates (Begum & Tso, 2016; Tso,
70 Miller, & Begum, 2017) – opposite to the amblyopia induced by monocular deprivation during the
71 critical period. The mechanism supporting the perceptual boost of the deprived information could
72 be either a form of homeostatic plasticity (like that observed in animal models), and/or a release of
73 contrast adaptation for the deprived stimulus (Blakemore & Campbell, 1969; Boynton et al., 1999;
74 Gardner et al., 2005; Maffei, Fiorentini, & Bisti, 1973; Movshon & Lennie, 1979). Irrespective of
75 the interpretation, the data clearly indicate that effects can be long-lasting or even permanent. For
76 example, in patients with keratoconus (adult-onset corneal dystrophy, often monocular), best
77 corrected visual acuity is worse than in emmetropic eyes, but it is better than predicted by the
78 corneal dystrophy (Sabesan & Yoon, 2009, 2010): when corneal aberrations of the keratoconic
79 (KC) eyes are simulated in the emmetropic eyes, visual acuity is worse than in the KC eyes,
80 demonstrating a permanent perceptual boost of the deprived information. Moreover, in adult
81 amblyopes (Lunghi et al., 2018), short-term monocular deprivation (of the amblyopic eye) may lead

82 to permanent partial recovery of acuity (of the amblyopic eye). This observation resonates with the
83 idea – introduced in the context of work at the cellular level – that homeostatic plasticity and
84 Hebbian plasticity may be fundamentally linked (Maffei & Turrigiano, 2008) and may open
85 important new pathways for the therapy of amblyopia and, in general, for the rehabilitation of early-
86 onset visual dysfunctions (Legge & Chung, 2016).

87 This possibility highlights the importance of understanding the neural substrates of short-term
88 deprivation in adult humans. So far, monocular deprivation effects have been indirectly studied with
89 MR spectroscopy (showing a GABA concentration change in the occipital cortex, Lunghi, Emir, et
90 al., 2015) and Visual Evoked Potentials (showing a modulation of the early visual response
91 components, Lunghi, Berchicci, et al., 2015). Indirect evidence also indicates that deprivation
92 effects are not generalized but preferentially involve the parvocellular pathway – given that effects
93 are more prominent and longer-lasting for chromatic equiluminant stimuli in humans (Lunghi, Burr,
94 & Morrone, 2013), and strongest in macaques when deprivation mainly affects the parvocellular
95 activity (Begum & Tso, 2016). Here we directly measure the changes in early visual cortical areas
96 using 7T fMRI in adult humans, before and after two hours of monocular deprivation. Assessing the
97 BOLD change and its selectivity to spatial frequency with a newly developed approach
98 (conceptually similar to the population Receptive Field method, Dumoulin & Wandell, 2008), we
99 demonstrate a change of ocular drive of BOLD signals in primary visual cortex, selective for the
100 higher spatial frequencies and strongest along the ventral pathway, consistent with a stronger
101 plasticity potential of the parvocellular pathway in adulthood.

102 Results

103 Monocular deprivation boosts V1 responses to the deprived eye and shifts BOLD ocular dominance

104 To investigate the visual modulation of BOLD signal by short term deprivation, we performed
105 ultra-high field (UHF, 7T) fMRI during the presentation of high contrast dynamic visual stimuli,
106 delivered separately to the two eyes, before and after 2h of monocular contrast deprivation (see
107 schematic diagram in Fig. 1A).

108 The reliability and high signal-to-noise ratio of our system allow us to obtain significant activations
109 with only two blocks of stimulation (Fig. 1C shows the profile of V1 BOLD response), thereby
110 targeting the first 10 minutes after deprivation, when the perceptual effects are strongest (Lunghi,
111 Burr, & Morrone, 2011; Lunghi, Burr, & Morrone, 2013). As shown in Fig. 1B, the stimulation was
112 sufficient to reliably activate most early visual areas (dashed lines outline ROIs limited by stimulus
113 eccentricity, as detailed in the methods).

114 We measured the plasticity effect by comparing activity before/after deprivation in response to
115 stimulation in the two eyes with low- and high-spatial frequency bandpass stimuli that differentially
116 stimulate the magno- and parvocellular pathways (see Figure 1 - figure supplement 1 panels C-D
117 for maps of responses to stimuli in both eyes, before and after deprivation). Consistent with prior
118 evidence suggesting higher susceptibility to plasticity of the parvocellular pathway (Lunghi,
119 Berchicci, et al., 2015; Lunghi, Burr, & Morrone, 2011; Lunghi, Emir, et al., 2015; Lunghi & Sale,
120 2015), we observe a strong effect of Monocular Deprivation on BOLD responses to stimuli of high
121 spatial frequency (peak 2.7 cycles per degree, high-frequency cut-off at half-height 7.5 cpd). Fig.
122 1D shows that the V1 response to the high spatial frequency stimuli presented in the left and right
123 eye is nearly equal before deprivation (“PRE”) (see Figure 1 - figure supplement 1, panels C-D and
124 Figure 1 - figure supplement 2, panel A, mapping the difference between responses to the two
125 eyes). However, after deprivation (“POST”), the response in the two eyes changes in opposite
126 directions, with a boost of the BOLD response (measured as GLM Beta values, expressed in units
127 of % signal change) of the deprived eye and a suppression of the non-deprived eye (see also Figure
128 1 - figure supplement 2, panel B). This was formally tested with a two-way repeated measure
129 ANOVA, entered with the mean BOLD responses across all vertices in the left and right V1 region,
130 for the four conditions and each participant (Fig. 1D show averages of this values across
131 participants). The result reveals a significant interaction between the factors *time* (PRE, POST
132 deprivation) and *eye* (deprived, non-deprived; interaction term $F(1,18) = 13.80703$, $p = 0.00158$; the
133 result survives a split-half reliability test: see Figure 1 - figure supplement 3).

134 *Figure 1: Monocular deprivation modulates 7T BOLD responses in early visual cortex*

135 Fig. 1E confirms these findings with an analysis of the aggregate subject data, obtained by pooling
136 all V1 vertices across all subjects. For each vertex, we defined an index of Ocular Dominance
137 computed as the difference of BOLD response to the deprived and non-deprived eye. This index is
138 not to be confused with the anatomical arrangement of vertices with different eye preference that
139 define the ocular dominance columns (Cheng, Waggoner, & Tanaka, 2001; Yacoub et al., 2007),
140 that cannot be directly imaged with voxel size of 1.5mm. However, at this low resolution, each
141 voxel is expected to average signals from a biased sample of ocular dominance columns leading to
142 an eye preference of that particular voxel (the Ocular Dominance index in Fig. 1E).

143 Before deprivation, the Ocular Dominance index is symmetrically distributed around zero,
144 indicating a balanced representation of the two eyes before deprivation (yellow distribution in
145 Fig.1E). After deprivation (black distribution in Fig.1E), the Ocular Dominance distribution shifts

146 to the right of 0, indicating a preference for the deprived eye (non-parametric Wilcoxon sign-rank
147 test comparing the PRE and POST Ocular Dominance medians, $z = 115.39$, $p < 0.001$).

148 In principle, the boost of responses to the deprived eye seen in Fig. 1D could be produced by
149 enhancing the response of vertices that originally preferred the deprived eye (without shifting ocular
150 dominance) or by changing Ocular Dominance of vertices that originally preferred the non-deprived
151 eye, driving them to prefer the deprived eye. The shift of the Ocular Dominance histogram in Fig.
152 1E is more compatible with the latter case, implying a recruitment of cortical resources for the
153 representation of the deprived eye. To investigate this further, we monitored the final POST-
154 deprivation Ocular Dominance of individual vertices that, PRE-deprivation, preferred the deprived
155 eye (yellow half distribution in Fig 2B). The majority of vertices continue to prefer the same eye
156 before and after deprivation. The median Ocular Dominance is significantly larger than 0 both PRE
157 and POST (Wilcoxon sign-rank test, $z > 101.54$, $p < 0.0001$ in both cases) and the correlation
158 between Ocular Dominance indices before and after deprivation is strong and positive (Pearson's
159 $R(32236) = 0.22$ [0.21-0.23], $p < 0.0001$). Note that a completely random reassignment of Ocular
160 Dominance after deprivation would have produced a histogram centered at 0 and no correlation
161 between Ocular Dominance indices PRE- and POST deprivation. This is not consistent with the
162 results of Fig. 2B, which thereby provide evidence that our estimates of Ocular Dominance before
163 and after deprivation are congruent, even though they were collected in different fMRI sessions
164 separated by 2h. In addition, the distribution of Ocular Dominance after deprivation is well
165 predicted by adding only a small amount of noise to the original half distribution (Gaussian noise
166 with 0.12 standard deviation, black line), suggesting that these vertices were largely unaffected by
167 monocular deprivation. This is also supported by the repeated measure ANOVA of individual
168 subject data (Fig. 2A), revealing a strong main effect of eye ($F(1,18) = 48.28901$, $p < 10^{-5}$): the
169 response to the deprived eye is stronger than the non-deprived eye, both before deprivation (due the
170 selection, $t(18) = -8.616$, $p < 10^{-5}$), and after deprivation ($t(18) = -4.281$, $p < 10^{-5}$), with no effect of
171 time and no $time \times eye$ interaction (all $F(1,18) = 0.20429$, $p > 0.5$).

172 A completely different pattern is observed for the vertices originally preferring the non-deprived
173 (yellow half-distribution in Fig. 2D). Here the distribution of Ocular Dominance clearly shifts after
174 deprivation; the median moves from significantly negative before deprivation (Wilcoxon sign-rank
175 test, $z = -175.97$, $p < 0.0001$) to significantly positive after deprivation (Wilcoxon sign-rank test,
176 $z = 64.46$, $p < 0.0001$), implying a shift of dominance in favor of the deprived eye. Again, this is not
177 consistent with a random reassignment of Ocular Dominance after deprivation, which predicts a
178 distribution centered at 0. Contrary to Fig. 2B, the POST- Ocular Dominance distribution cannot be

179 predicted by injecting Gaussian noise to the PRE- Ocular Dominance distribution (black line, 0.12
180 standard deviation like for Fig. 2B): for these vertices, there is a shift of Ocular Dominance with
181 short term monocular deprivation. This is confirmed with the repeated measure ANOVA (Fig. 2C),
182 where the $time \times eye$ interaction is significant ($F(1,18) = 44.82812$, $p < 10^{-5}$), implying a different
183 modulation PRE and POST deprivation. In addition and crucially, POST-deprivation BOLD
184 responses to the deprived eye are significantly larger than POST-deprivation responses to the non-
185 deprived eye ($t(18) = -2.775$ $p = 0.012$; whereas, by selection, the opposite is true before
186 deprivation: $t(18) = 12.034$, $p < 10^{-5}$).

187 *Figure 2: Monocular deprivation shifts 7T BOLD Ocular Dominance in V1*

188 In summary, Ocular Dominance before deprivation defines two similarly sized sub-regions of V1
189 vertices ($44.58 \pm 5.38\%$ and $55.42 \pm 5.38\%$ of analyzed V1 vertices; $44.84 \pm 5.12\%$ and $55.16 \pm$
190 5.12% of all V1 vertices) with radically different behaviors that are not consistent with an artifact
191 induced by vertex selection. The sub-region that originally represents the deprived eye does not
192 change with deprivation; the sub-region that originally represents the non-deprived eye is
193 rearranged with deprivation, as a large portion of vertices turn to prefer the deprived eye.

194 If plasticity were not eye-specific and/or we failed to match our V1 vertices before/after
195 deprivation, we would expect that splitting the distribution of V1 ocular dominance generates
196 opposite effects in the two subpopulations: vertices preferring the deprived eye before deprivation
197 should swap to prefer the other eye, mirroring the effect seen in the vertices preferring non-deprived
198 eye. This is not seen, implying that we did successfully match vertices across the 2h of deprivation
199 and that the selective Ocular Dominance shift, observed for about half of our vertices, is not an
200 artifact.

201 We also measured the perceptual effects of short-term monocular deprivation effects using
202 Binocular Rivalry, just before the PRE- and POST-deprivation fMRI sessions. In line with previous
203 studies (Binda & Lunghi, 2017; Lunghi, Berchicci, et al., 2015; Lunghi, Burr, & Morrone, 2011;
204 Lunghi, Emir, et al., 2015; Lunghi & Sale, 2015), short-term monocular contrast deprivation
205 induced a 30% increase of phase duration for the deprived eye (POST to PRE-deprivation ratio:
206 1.31 ± 0.30) and a 15% decrease of phase duration for the non-deprived eye (ratio: 0.86 ± 0.30),
207 producing a significant $time \times eye$ interaction (Fig. 3A, repeated measure ANOVA on the mean
208 phase durations for each participant, interaction: $F(1,18) = 23.56957$, $p = 0.00013$). This effect size
209 is similar to that measured in recent experiments using the same paradigm, but letting subjects
210 continue normal activity during the 2h of monocular deprivation (Lunghi, Burr, & Morrone, 2011;

211 Lunghi, Emir, et al., 2015; Lunghi & Sale, 2015). This indicates that the prolonged high contrast
212 stimulation delivered for retinotopic mapping to the non-deprived eye during the first ~30 minutes
213 of deprivation did not modulate the deprivation effects.

214 We defined a psychophysical index of the deprivation effect (DI_{psycho}) by using Eq. 6 in methods
215 section, where the POST to PRE-deprivation ratio of phase durations for the deprived eye, is
216 divided by the same ratio for the non-deprived eye. Values larger than 1 imply a relative increase of
217 the deprived eye phase duration, i.e. the expected effect; a value less than 1 indicates the opposite
218 effect and a value of 1 indicates no change of mean phase duration across eyes. All but two subjects
219 have values larger than 1, indicating a strong effect of deprivation. However, the scatter is large
220 with values ranging from 0.7 to 3, suggesting that susceptibility to visual plasticity varies largely in
221 our pool of participants. Capitalizing on this variability, we tested whether the size of the
222 psychophysical effect correlates with the BOLD effect across participants. Using the same Eq. 6 to
223 compute the deprivation effect on BOLD responses (DI_{BOLD}), we observed a strong correlation
224 between the effect of monocular deprivation on psychophysics and BOLD (shown in Fig. 3B).
225 Subjects who showed a strong deprivation effect at psychophysics ($DI_{\text{psycho}} > 2$) also showed a
226 strong deprivation effect in BOLD responses ($DI_{\text{BOLD}} = 1.85 \pm 0.42$). Given that the psychophysics
227 was measured only for central vision and at 2 cpd stationary grating, whereas BOLD responses
228 were pooled across a large portion on V1 and were elicited using broadband dynamic stimuli, the
229 correlation suggests that the psychophysical effect may be used as a reliable proxy of a general
230 change of cortical excitability, which can be measured by fMRI.

231 *Figure 3: Deprivation effects on BOLD and on psychophysics are correlated*

232 *Monocular deprivation shifts BOLD Spatial Frequency Tuning for the deprived eye*

233 The BOLD measure we use here gives us the chance to measure the effect of Monocular
234 Deprivation across spatial frequencies and as function of eccentricity. We used 5 band-pass noise
235 (1.25 octaves half-width at half maximum) stimuli with peak spatial frequency selected to have a
236 complete coverage of spatial frequencies from 0.03 to 12.5 cpd (see Figure 4 - figure supplement 1).
237 In contrast with the strong and reliable plasticity of responses to the high spatial frequency stimulus
238 (peaking at 2.7 cpd, Figs. 1-3), we find that the plasticity effect is absent at low spatial frequencies
239 (interaction index for the highest spatial frequency stimulus: 0.70 ± 0.19 ; for the lowest spatial
240 frequency stimulus: 0.16 ± 0.15 ; paired t-test $t(18) = -3.441$, $p = 0.003$).

241

242 *Figure 4: Deprivation affects spatial frequency selectivity in V1*

243 Thus, monocular deprivation produces a change of the spatial frequency selectivity of the V1
244 BOLD response. Before deprivation, the BOLD response shows a broad band-pass selectivity for
245 our stimuli, with a preference for the stimulus peaking at intermediate spatial frequencies, between
246 0.4 and 1.1 cpd, and a slight attenuation at higher spatial frequencies, similar for the two eyes (Fig.
247 4A). After deprivation (Fig. 4B), the non-deprived eye shows similar selectivity and an overall
248 decrease of responses. For the deprived eye, the shape of the curve changes: from band-pass to
249 high-pass, implying that the enhancement affects primarily the higher spatial frequencies.

250 To model this effect, we assume that each vertex on the cortical surface subtends a multitude of
251 neuronal channels, each with narrow tuning for spatial frequency and collectively spanning a large
252 range of spatial frequencies – an approach conceptually similar to the population Receptive Field
253 model for retinotopic mapping (Dumoulin & Wandell, 2008). Independently of the exact bandwidth
254 and peak preference of the neuronal population contributing to the final BOLD selectivity, we find
255 that the shape of all these curves is captured with a simple one-parameter model: what we term the
256 population tuning for Spatial Frequency. This is given by a Difference-of-Gaussians (DoG) function
257 with one free parameter, the spatial constant (while the excitation/inhibition spatial constant ratio is
258 fixed; see eq. 4 in the Methods and curves in Figure 5 - figure supplement 1). The free parameter
259 sets the high spatial frequency cut-off at half-height of the filter. The continuous lines in Fig. 4
260 show how the model fits the grand-average of V1 responses, with best fit cut-off around 5 cpd
261 similar for all conditions except for the POST-deprivation deprived eye, where the cut-off is 6.2 cpd
262 (single vertex examples are given in Figure 5 - figure supplement 1 panels C-I). The DoG equation
263 has been successfully used in previous studies to model CSF and neural responses at variable
264 stimulus parameters e.g. illumination levels (Enroth-Cugell & Robson, 1966; Hawken, Parker, &
265 Lund, 1988), validating this equation for modeling the overall selectivity of large neuronal
266 ensembles.

267 *Figure 5: population Spatial Frequency Tuning in V1*

268 Using this model to analyze single vertex responses, we evaluated the best-fit spatial frequency cut-
269 off of the neural population contributing to the vertex BOLD response (see details in the methods
270 and Figure 5 - figure supplement 1 panels A-C; briefly, we used the DoG model to predict the
271 response elicited by our five band-pass noise stimuli in populations with different spatial frequency
272 selectivity, i.e. filters with different cut-off; we then found the cut-off value that maximizes the
273 correlation between the predicted responses and the observed BOLD responses). We used this
274 procedure to fit BOLD responses in each of our four conditions, estimating spatial frequency

selectivity in individual vertices in each condition: separately for the two eyes, PRE/POST deprivation. Before deprivation, the spatial frequency cut-off decays with eccentricity as expected. Fig 5A maps both eccentricity (pRF eccentricity estimates from a separate retinotopic mapping scan) and spatial frequency cut-off values, obtained by fitting responses to the deprived eye, before deprivation (averaged across hemispheres and subjects). The cut-off is around 16 in the para-fovea (eccentricity around 1.5 deg) and down to 4 in the periphery (eccentricity around 8 deg). This relationship between eccentricity and spatial frequency preference is consistent with previous fMRI results (D'Souza et al., 2016; Henriksson et al., 2008) and with psychophysics (Rovamo, Virsu, & Nasanen, 1978). The model captures well the selectivity of an example V1 vertex (Fig. 5B, goodness of fit better than 0.9), sampled from the mid-periphery (3.4 deg) for the deprived eye, both before and after deprivation. The spatial frequency cut-off after deprivation shifts to higher values, increasing (in this example) by about a factor of three. Fig. 5C-D shows that this behavior is systematically observed across V1 vertices, but only for the deprived eye. Here the average cut-off is plotted as function of eccentricity, and the roll-off is consistent with the map in Fig. 5A. For the non-deprived eye, there is no effect of deprivation on spatial frequency selectivity (Fig. 5C). In contrast, for the deprived eye (Fig. 5D), there is a shift towards preferring higher spatial frequencies, at all eccentricities, which is captured by an increased value of the cut-off frequency parameter leading to an increased acuity of the BOLD response to the deprived eye.

Note that the change of spatial frequency selectivity for the deprived eye is most evident at eccentricities of 4 deg and higher (see Fig. 5D), where vertices have peak sensitivity at mid-to-low spatial frequencies before deprivation. In the fovea, where many vertices already prefer the highest spatial frequency stimulus before deprivation, our fitting procedure is likely to underestimate the change of spatial frequency selectivity. Importantly, the spatial frequency selectivity for the non-deprived eye is unchanged at all eccentricities, corroborating the eye and stimulus specificity of the short-term monocular deprivation effect. These findings are consistent with maps in Figure 1 - figure supplement 1 panels C-D showing that deprivation effects are largely homogenous across all V1 eccentricities, with no obvious clustering of effects in the fovea or in the periphery.

Figure 6: Deprivation effects on the deprived eye population Spatial Frequency Tuning and binocular rivalry phase duration are correlated

To test the significance of these effects, we pooled the best fit cut-off values from all selected V1 vertices across eccentricities and averaged them across participants (Fig. 6A). The repeated measure ANOVA (performed on the log-transformed values, which are distributed normally as assessed by the Jarque-Bera test) shows no significant $time \times eye$ interaction ($F(1,18) = 3.67607$, $p = 0.07121$)

and non significant main effect of time ($F(1,18) = 2.62546$, $p = 0.12255$) but a significant main effect of eye ($F(1,18) = 13.58079$, $p = 0.00169$). This is clarified by post-hoc t-tests revealing that the increase of spatial frequency cut-off for the deprived eye is significant ($t(18) = -2.263$, $p = 0.036$) whereas there is no significant change for the non-deprived eye ($t(18) = 0.440$, $p = 0.665$). Given that the *time* $\times$ *eye* interaction in the full V1 region is not significant, and to minimize noise contamination, we evaluated the effect of deprivation on spatial frequency cut-off at the individual level by a “Deprived Eye Change ($DepC_{cutoff}$)” index (Eq.7 in the methods), i.e. taking the POST vs. PRE-deprivation ratio of the spatial frequency cut-off for the deprived eye alone. As this ratio varies widely across participants, over more than 3 octaves, we asked whether this variability correlates with our psychophysical probe of plasticity: binocular rivalry. We used the same Eq. 7 to index the psychophysical change of the deprived eye ($DepC_{psycho}$), the POST to PRE- ratio of mean phase duration for the deprived eye, and found a strong positive correlation (Fig. 6B). POST-deprivation, the deprived eye shows an increase of mean phase duration (in binocular rivalry) and an increase of the spatial frequency cut-off (best fit of the BOLD responses): participants showing a stronger increase of phase duration, also showed a larger shift of selectivity towards higher spatial frequency. The correlation is consistent with the result of Fig. 3 showing that the enhancement of BOLD responses is correlated with the change of binocular rivalry and selective for the highest spatial frequency stimulus.

Monocular deprivation affects BOLD responses in the ventral stream areas beyond V1

We measured the effect over the main extra-striate visual cortical areas. The selective boost of the deprived eye response to the high spatial frequency is as strong in V2 as in V1 (Fig. 1 - figure supplement 1 and Fig. 7E). The boost is present also in V3 and V4. In V4 the boost appears to be present also for lower spatial frequencies, but again only for the deprived eye (Fig. 7A-B), possibly reflecting the larger spatial frequency bandwidth of V4 neurons compared to V1.

Figure 7: Deprivation effects are stronger in ventral than in dorsal stream areas.

The results are very different for dorsal area V3a (Fig. 7C-D) and hMT+ (Fig. 7E-F), which do not show any significant change of responses in either eye at high spatial frequencies. Although the preferred response moves to lower spatial frequencies, consistent with a stronger input of the magnocellular pathway to the dorsal visual stream (Henriksson et al., 2008; Singh, Smith, & Greenlee, 2000), the response to the highest spatial frequency stimulus is still strong and reliable in both V3a and hMT+. Note that the reliable BOLD estimates of Fig. 7 are computed after pooling vertices within the ROI and then averaging across subjects. However, the response of hMT+

340 evaluated at the individual vertex do not show significant activation (Fig. 1B), probably reflecting
341 more variable organization of activity within this ROI across subjects (Smith et al., 2006).

342 Fig. 7G quantifies the effect of short-term monocular deprivation (using the ANOVA time x eye
343 interaction term, which measures the eye-selective modulation of BOLD response after deprivation
344 for the highest spatial frequency) across the main visual areas. The plasticity effect is strongest in
345 V1, V2 and V3; it is still strong and significant in ventral area V4 ($t(18) = 2.41$ $p = 0.0270$), but it is
346 absent in V3a and hMT+, where the time x eye interaction is not significantly different from 0
347 ($t(18) = 0.52$ $p = 0.6115$ and $t(18) = -0.19$ $p = 0.8513$ respectively). The plasticity effect in ventral
348 area V4 is significantly stronger than in dorsal areas V3a and hMT+ ($t(18) = 2.39$, $p = 0.0278$ and
349 $t(18) = 2.36$, $p = 0.0299$ for V4-V3a and V4-hMT+ respectively).

350 This result suggests a preferential involvement of the parvocellular vs. magnocellular pathway,
351 leading to the differential plasticity effect in extra-striate visual areas of the ventral and dorsal
352 pathway. Interestingly, the plasticity effect is robust in areas where the majority of cells are
353 binocular (like V3 and V4), indicating that the effect does not require segregated representations of
354 the two eyes (e.g. ocular dominance columns).

355 Discussion

356 We demonstrate that two hours of abnormal visual experience has a profound impact on the neural
357 sensitivity and selectivity of V1. BOLD activity across the V1 cortical region paradoxically
358 increases for the eye that was deprived of contrast vision, and decreases for the eye exposed to
359 normal visual experience.

360 The enhanced response to the deprived eye fits well with the concept of homeostatic plasticity, first
361 observed in rodent visual cortex, both juvenile and adult (Maffei, Nelson, & Turrigiano, 2004;
362 Mrsic-Flogel et al., 2007; Turrigiano & Nelson, 2004), which is the tendency of neural circuits to
363 keep the average firing rates constant in spite of anomalous stimulation (Maffei & Turrigiano, 2008;
364 Turrigiano, 2012) (Mrsic-Flogel et al., 2007; Turrigiano & Nelson, 2004). More recently, similar
365 observations have been made in the adult macaque V1 after two hours of monocular deprivation
366 during anesthesia (Begum & Tso, 2016; Tso, Miller, & Begum, 2017). The post-deprivation gain
367 boost observed in the monkey is consistent with our observations of an increased BOLD response to
368 the deprived eye. We also observe an antagonistic suppression of the non-deprived eye BOLD
369 response; together, the two effects lead to a shift of ocular preference of individual vertices in favor
370 of the deprived eye. However, this effect is only observed in those V1 vertices that responded

371 preferentially to the non-deprived eye before deprivation. No change of ocular preference is seen in
372 vertices that already prefer the deprived eye before deprivation, which maintain their eye-preference
373 after deprivation. This pattern of results cannot be explained by an overall gain increase; rather, it is
374 consistent with the idea that the representation of the deprived eye recruits cortical resources (which
375 may or may not correspond to cortical territory), normally dedicated to the other eye.

376 A similar antagonist effect on the two eyes (boosting the deprived eye and suppressing the non-
377 deprived eye) was also observed in the VEP responses after short-term monocular deprivation
378 (Lunghi, Berchicci, et al., 2015), and could be implemented through a modulation of the
379 excitatory/inhibitory circuitry. Regulation of the excitation/inhibition balance through GABAergic
380 signaling is considered to be a key factor for cortical plasticity, including homeostatic plasticity
381 (Maffei & Turrigiano, 2008). Interestingly, the involvement of GABAergic signaling in the effect
382 of short-term monocular deprivation is directly supported by MR Spectroscopy data in adult
383 humans, showing that resting GABA in a large region of the occipital cortex is specifically reduced
384 after short-term monocular deprivation (Lunghi, Emir, et al., 2015).

385 The functional relevance of the BOLD changes we observe is demonstrated by their correlation
386 with our behavioral assay of plasticity, obtained through binocular rivalry. This correlates both with
387 the BOLD ocular dominance change (relative boost/suppression of the deprived/non-deprived eye),
388 and with the BOLD acuity change for the deprived eye (change of spatial frequency tuning,
389 assessed with our pRF-like modeling approach). The correlation holds despite binocular rivalry
390 being restricted to foveal vision, whereas the assessment of BOLD plasticity is pooled across V1
391 (including the mid-periphery). This implies that the change of binocular rivalry dynamics is a proxy
392 for the more general plasticity effects that involves the whole primary visual cortex. This finding
393 has long reaching implications, as it could validate the use of binocular rivalry as a biomarker of
394 adult cortical plasticity, based on the neural mechanisms revealed by the present 7T fMRI results.
395 Interestingly, the binocular rivalry phenomenon originates in the primary visual cortex – probably
396 at the earliest stages – and is an expression of the dynamics of excitatory transmission and
397 inhibitory feedback (Tong, Meng, & Blake, 2006); as such it is a measure that could reflect the
398 overall excitation-inhibition ratio (van Loon et al., 2013), and its modulation in plasticity (Lunghi,
399 Emir, et al., 2015; Maffei & Turrigiano, 2008).

400 Our data support the notion that V1 circuitry may be optimized by perceptual experience (Fiorentini
401 & Berardi, 1980). They are also consistent with a large perceptual learning literature suggesting that
402 associative cortical areas retain a high degree of flexibility (Doshier & Lu, 2017; Doshier & Lu,

1999; Fuchs & Flugge, 2014; Harris, Gliksberg, & Sagi, 2012; Kahnt et al., 2011; Karni et al., 1995; Lewis et al., 2009; Shibata et al., 2012; Watanabe & Sasaki, 2015). Although the monocular deprivations effects observed here are more robust in V1, they are reliable in V2 and V3 as well. However, a clear difference emerges between extra-striate visual areas in the ventral and dorsal stream. While ventral area V4 shows a strong deprivation effect, area V3a, located at a similar tier in the dorsal stream, shows no BOLD change after short-term monocular deprivation. V4 is a primary target of the parvocellular system, which is best stimulated by our highest spatial frequency stimulus; V3a and hMT+ are preferential targets of the magnocellular system, which respond more strongly to our lower spatial frequency stimuli (see Fig. 7). The different plasticity response of the ventral and dorsal stream, together with the selectivity for the high spatial frequencies of the V1 plasticity, suggests that the parvocellular pathway is most strongly affected by short-term plasticity. This is consistent with the finding in non-human primates that deprivation of the stimuli that optimally drive the parvocellular system is sufficient to produce a reliable plasticity effect (Begum & Tso, 2016). It is also consistent with the finding that the effect of short-term monocular deprivation is strongest and more long-lasting for chromatic equiluminant stimuli (Lunghi, Burr, & Morrone, 2013) .

Other evidence shows that short-term deprivation may affect other properties of vision. In particular, selective deprivation of orientation (Zhang et al., 2009) or spatial frequency (Zhou, Reynaud, & Hess, 2014) or color (Zhou et al., 2017) or even simply phase scrambling of the image in one eye (Bai et al., 2017) may lead to a boost of the deprived signal. These effects have been interpreted as a form of release of inhibition from the adapted signal (Zhang et al., 2009) – a concept that is not distant from homeostatic plasticity, where the network aims to keep overall activity constant. The conceptual border between adaptation and plasticity is fuzzy, given that some mechanisms are shared and both effects have the same outcomes. Be it adaptation or plasticity, the monocular deprivation mechanisms are probably cortical and affect mainly the deprived eye. There is evidence that the boost of the deprived eye is also observed when the two eyes receive equally strong stimulation, but perception of one eye stimulus is suppressed experimentally (by the continuous flash suppression technique Kim, Kim, & Blake, 2017); this result dismisses the retinal or thalamic contribution to the deprivation effect. Only in rare occasions does adaptation induce effects that last over days (McCollough, 1965), yet our recent work shows that deprivation effects of short-term monocular deprivation is retained across 6h sleep (Menicucci et al., 2018), consistent with plasticity reinforcement during sleep (Raven et al., 2018; Timofeev & Chauvette, 2017). Most importantly, in adult amblyopic patients, short-term monocular deprivation is able to induce improvement of visual acuity and stereovision (Lunghi et al., 2018) for up to one year. All this

evidence supports the concept that homeostatic plasticity in the human adult cortex may be linked with or may promote more stable forms of Hebbian-like plasticity. This may endorse stable changes even in the adult brain, well after the closure of the critical period. Functional changes in associative cortex in adults have been demonstrated by short-term paired TMS studies (Chao et al., 2015). Interestingly, the decay of this functional connectivity change has a similar time-course as the monocular deprivation effect, about one hour. Also, Hebbian changes at the single cell level can be observed in V1 of adult anaesthetized cat, following activity pairing over a similar time-scale (from minutes to a few hours) (Fregnac et al., 1988). All these results demonstrate that V1 retains potential for Hebbian plasticity outside the critical period – although it may need particular conditions to exploit such potential.

Understanding homeostatic plasticity and its relation to Hebbian plasticity may be fundamental to open the way to new approaches to treat brain dysfunction. Particularly important is ocular dominance plasticity in amblyopia (Webber & Wood, 2005), a cortical deficit still without cure in adults, although recent advancements in training procedures are opening new hopes (Levi & Li, 2009; Sengpiel, 2014). Endorsing plasticity may increase the effectiveness of these treatments and preliminary data from our laboratory suggest that monocular deprivation of the amblyopic eye may indeed boost sensitivity of the deprived eye and improve its acuity (Lunghi et al., 2018) – just like an acuity change is revealed by the present BOLD measurements in normally sighted participants. Our data demonstrate that two hours of abnormally unbalanced visual experience is sufficient to induce a functional reorganization of cortical circuits, particularly of the parvocellular pathway, leading to an alteration of basic visual perceptual abilities.

[Methods text](#)

[Key resource table](#)

Reagent type (species) or resource	Designation	Source or reference	Identifiers	Additional information
software, algorithm	Freesurfer v6.0.0	(Fischl et al., 2002)	SCR_001847	
software, algorithm	SPM	(Friston, 2007)	SCR_007037	
software, algorithm	BrainVoyager	(Goebel, Esposito, & Formisano, 2006)	SCR_006660	
software, algorithm	FSL	(Jenkinson et al., 2012)	SCR_002823	
software, algorithm	MATLAB	MathWorks	SCR_001622	
software, algorithm	PsychToolbox	(Brainard, 1997)	SCR_002881	

460

461 EXPERIMENTAL MODEL AND SUBJECT DETAILS

462 *Human subjects*

463 Experimental procedures are in line with the declaration of Helsinki and were approved by the
464 regional ethics committee [Comitato Etico Pediatrico Regionale—Azienda Ospedaliero-
465 Universitaria Meyer—Firenze (FI)] and by the Italian Ministry of Health, under the protocol
466 “Plasticità e multimodalità delle prime aree visive: studio in risonanza magnetica a campo ultra alto
467 (7T)” version #1 dated 11/11/2015. Written informed consent was obtained from each participant,
468 which included consent to process and preserve the data and publish them in anonymous form.

469 Twenty healthy volunteers with normal or corrected-to-normal visual acuity were examined (8
470 females and 12 males, mean age = 27 years).

471 METHOD DETAILS

472 *Experimental design*

473 Each participant underwent two scanning sessions separated by two hours, during which they were
474 subject to the short-term monocular deprivation procedure described below. Just before each
475 scanning session, their binocular rivalry was measured psychophysically. One (male) participant
476 was excluded because of strong eye dominance tested with binocular rivalry before the deprivation.
477 This left 19 participants (8 females and 11 males) whose complete datasets were entered all
478 analyses. Sample size was set to enable testing for correlations between neuroimaging and
479 psychophysical data. Previous work (Lunghi, Emir, et al., 2015) reveals a correlation between MR
480 spectroscopy data and binocular rivalry measures $r = 0.62$ (or higher), which implies a minimum of
481 17 participants to detect a significant correlation at 0.05 significance level, with test power of 80%
482 (Lachin, 1981).

483

484 *Short-term Monocular Deprivation*

485 Monocular deprivation was achieved by patching the dominant eye for 2 hours. The operational
486 definition of dominant eye applied to the eye showing the longer phase durations during the
487 baseline binocular rivalry measurements. Like in previous studies (Binda & Lunghi, 2017; Lunghi,
488 Burr, & Morrone, 2011; Lunghi, Burr, & Morrone, 2013), we used a translucent eye-patch made of
489 plastic material allowing light to reach the retina (attenuation 0.07 logUnits, at least 3 times smaller
490 than the threshold for discriminating a full-field luminance decrement (Knau, 2000) and more than
491 ten times smaller than the minimum photopic luminance decrement required for shifting the spatial
492 (Van Nes & Bouman, 1967) or temporal contrast sensitivity function (Kelly, 1961)). The patch

493 prevents pattern vision, as assessed by the Fourier transform of a natural world image seen through
494 the eye-patch. During the 2 hours of monocular deprivation, observers were either engaged in the
495 retinotopic mapping experiment (about 30', described below) or they were free to read and use a
496 computer.

497 *Binocular Rivalry*

498 Binocular rivalry was measured in two short sessions (each comprising two runs of 3 minutes each),
499 immediately before the Pre- and Post-deprivation MR sessions, in a quiet space adjacent to the MR
500 control room. Visual stimuli were created in MATLAB running on a laptop (Dell) using
501 PsychToolbox (Brainard, 1997), and displayed on a 15- inch monitor (BenQ). Like in (Lunghi,
502 Emir, et al., 2015), observers viewed the visual stimuli presented on the monitor at a distance of 57
503 cm through anaglyph red-blue goggles (right lens blue, left lens red). Responses were recorded with
504 the computer keyboard by continuous alternate keypresses. Visual stimuli were two oblique
505 orthogonal red and blue gratings (orientation: $\pm 45^\circ$, size: 3° , spatial frequency: 2 cpd, contrast
506 50%), surrounded by a white smoothed circle, presented on a black uniform background in central
507 vision. Peak luminance of the red grating was reduced to match the peak luminance of the blue one
508 using photometric measures. All included participants had typical binocular rivalry dynamics, with
509 low percentage of mixed percepts (reported for $8.5 \pm 2.04\%$ of time on average). Only one
510 participant experienced of mixed percepts for more than 20% of time (exactly for 31.2%) and his
511 data are in line with the others'.

512 *Stimuli for fMRI*

513 Visual stimuli were projected with an MR-compatible goggle set (VisuaStimDigital, Resonance
514 Technologies, Los Angeles, USA), connected to a computer placed in the control room. The
515 goggles covered a visual field of approximately 32×24 deg, with a resolution of 800×600 pixels,
516 mean luminance 25 cd/m^2 ; the images in the two eyes were controlled independently.

517 During all functional MRI scans participants were instructed to maintain central fixation on a red
518 point (0.5 degrees) that was constantly visible at the center of the screen. Bandpass noise stimuli
519 were white noise images filtered to match the spatial frequency tuning of neurons in the visual
520 cortex (Morrone & Burr, 1988). We generated a large white noise matrix (8000×6000) and filtered
521 it with a two-dimensional circular bandpass filter Bp defined by Eq. 1:

$$522 \quad Bp = e^{\frac{-\ln(\frac{w}{P})^2}{2[q*\ln(2)]^2}} \quad \text{eq. 1}$$

523 where P is the peak spatial frequency, q is the filter half-width at half maximum in octaves. We
524 generated five band-pass noise stimuli, by setting $q = 1.25$ octaves and $P = 0.1$ cpd, 0.2 cpd, 0.4
525 cpd, 1.1 cpd, 2.7 cpd. Each stimulus was presented for a block of 3TRs, during which the image
526 was refreshed at 8Hz (randomly resampling a 800×600 window from the original matrix). Stimuli
527 were scaled to exploit the luminance range of the display, and this yielded very similar RMS
528 contrast values (shown in Figure 4 - figure supplement 1). Stimulus blocks were separated by 4TRs
529 blanks, consisting of a mid-level gray screen. The five band-pass noise stimuli blocks were
530 presented in pseudo-random order, twice per run, for a total of 70 TRs. In each run, stimuli were
531 only presented to one eye, while the other was shown a mid-level gray screen. Each eye was tested
532 once, before and after deprivation.

533 Immediately upon application of the monocular patch, we performed two additional scans to
534 perform retinotopic mapping of visual areas. Meridian and ring stimuli were presented monocularly
535 (to the non-patched eye) and were defined as apertures of a mid-level gray mask that uncovered a
536 checkerboard pattern, 1 deg at 1 deg eccentricity to 2.5 deg at 9 deg eccentricity, rotating and
537 contracting at a rate of one check per second. Meridians were defined by two 45° wedges centered
538 around 0° or around 90° . The horizontal and vertical meridian were presented interchangeably for 5
539 TRs each (without blanks) and the sequence was repeated 6 times for a total of 60 TRs. Rings
540 partitioned screen space into six contiguous eccentricity bands (0-0.9 deg, 0.9-1.8 deg, 1.8-3.3 deg,
541 3.3-4.7 deg, 4.7-6.48 deg, 6.48-9 deg). Odd and even rings were presented in two separate runs. In
542 each run, the three selected rings and one blank were presented in random order for 5 TRs each, and
543 the sequence was repeated (with different order) 6 times for a total of 120 TRs.

544 *MR system and sequences*

545 Scanning was performed on a Discovery MR950 7 T whole body MRI system (GE Healthcare,
546 Milwaukee, WI, USA) equipped with a 2-channel transmit driven in quadrature mode, a 32-channel
547 receive coil (Nova Medical, Wilmington, MA, USA) and a high-performance gradient system (50
548 mT/m maximum amplitude and 200 mT/m/ms slew rate).

549 Anatomical images were acquired at 1 mm isotropic resolution using a T1-weighted magnetization-
550 prepared fast Fast Spoiled Gradient Echo (FSPGR) with the following parameters: TR = 6 ms, TE =
551 2.2 ms, FA=12 deg, rBW = 50kHz, TI = 450 ms, ASSET = 2.

552 Functional images were acquired with spatial resolution 1.5 mm and slice thickness 1.4 mm with
553 slice spacing = 0.1 mm, TR = 3 ms, TE = 23ms, rBW = 250 kHz, ASSET = 2, phase encoding

554 direction AP-PA. No resampling was performed during the reconstruction. For each EPI sequence,
555 we acquired 2 additional volumes with the reversed phase encoding direction.

556 QUANTIFICATION AND STATISTICAL ANALYSIS

557 *ROI definition*

558 Areas V1, V2 and V3 were manually outlined for all participants using retinotopic data projected on
559 surface models of white matter. The V1/V2 boundary was traced from the vertical/horizontal
560 meridian flip superior/inferior to the calcarine sulcus, and the V2/V3 border and V3 border from the
561 subsequent opposite flips. Areas V4, V3a and hMT+ (merging MT and MST) were defined based
562 on the cortical parcellation atlas by Glasser et al. (Glasser et al., 2016). V1, V2, V3, V4 and V3a
563 ROIs were further restricted to select the representation of our screen space. Specifically, the
564 anterior boundaries were defined based on activation from most peripheral (6.48°-9°) ring stimuli of
565 the retinotopic mapping scans; in addition, vertices were only included in the analysis if their
566 preferred eccentricity (estimated through Population Receptive Field modelling, see below) was
567 larger than 1, since no reliable mapping could be obtained for the central-most part of the visual
568 field.

569 *Pre-processing of imaging data*

570 Analyses were performed mainly with Freesurfer v6.0.0, with some contributions of the SPM12 and
571 BrainVoyager 20.6 and FSL version 5.0.10 (Jenkinson et al., 2012) packages.

572 Anatomical images were corrected for intensity bias using SPM12 (Friston, 2007) and processed by
573 a standard procedure for segmentation implemented in Freesurfer (recon-all: Fischl et al., 2002). In
574 addition, each hemisphere was aligned to a left/right symmetric template hemisphere
575 (fsaverage_sym: Greve et al., 2013).

576 Functional images were corrected for subject movements (Goebel, Esposito, & Formisano, 2006)
577 and undistorted using EPI images with reversed phase encoding direction (Brain Voyager COPE
578 plug-in Jezzard & Balaban, 1995). We then exported the preprocessed images from BrainVoyager
579 to NiFTi format. These were aligned to each participant's anatomical image using a boundary based
580 registration algorithm (Freesurfer *bbergister* function) and projected to the cortical surface of each
581 hemisphere. All analyses were conducted on data in the individual subject space. In addition, for
582 visualization purposes, we also aligned the results of timecourse analyses (GLM and subsequent
583 pRF and spatial frequency tuning estimates) to the left/right symmetric template hemisphere.

584 Averaged results across the 18x2 hemispheres are shown in the maps of Fig. 1B, Fig. 5A and Figure
585 1 - figure supplement 1.

586 *GLM analysis of fMRI data*

587 General Linear Model analysis was performed with in-house MATLAB software (Mathworks,
588 version R2016b). We assumed that fMRI timecourses result from the linear combination of N
589 predictors: boxcar functions representing stimulus presence/absence (one per stimulus type)
590 convolved by a standard hemodynamic response function (see Eq. 2), plus two nuisance variables (a
591 linear trend and a constant). We modeled the hemodynamic response function as a gamma function
592 $h(t)$:

$$593 \quad h(t) = \frac{\left(\frac{t-\delta}{\tau}\right)^{(n-1)} e^{-\left(\frac{t-\delta}{\tau}\right)}}{\tau(n-1)!} \quad \text{eq. 2}$$

594 with parameters $n=3$, $t=1.5$ s, and $d=2.25$ s (Boynton et al., 1996). Beta weights of the stimuli
595 predictors were taken as estimates of the BOLD response amplitude and normalized by the
596 predictor amplitude to obtain a measure that directly corresponds to % signal change; beta weights
597 were also scaled by an error measure to obtain t-values, following the same procedure as in (Friston
598 et al., 1994). Computing BOLD responses for each individual vertex of the cortical surface leads to
599 up-sampling the functional data (each $1.5 \times 1.5 \times 1.5$ mm functional voxel projecting on an average
600 of 3 vertices). We ensured that this does not affect our statistical analyses by first averaging data
601 from all vertices within a region of interest (e.g. V1), thereby entering all ANOVAs with a single
602 value per subject and region of interest.

603

604 *Population Receptive Field mapping*

605 The pRFs of the selected voxels were estimated with custom software in Matlab, implementing a
606 method related to that described by Dumoulin and Wandell (Dumoulin & Wandell, 2008) and
607 provided as supplementary material. We modeled the pRF with a 1D Gaussian function defined
608 over eccentricity, with parameters ε and σ as mean and standard deviation respectively, and
609 representing the aggregate receptive field of all neurons imaged within the vertex area. We defined
610 the stimulus as a binary matrix S representing the presence of visual stimulation over space (here,
611 eccentricity between 0 and 10 deg with 40 steps per deg) for each of 6 ring stimuli. We used the
612 results of our GLM analysis to estimate the vertex response to each of our 6 rings (as t-values; using
613 beta values yields very similar results). We assumed that each vertex response is the linear sum

over space (eccentricity) of the overlap between the pRF of the voxel and the input stimulus, which is mathematically equivalent to the matrix multiplication between the stimulus and the pRF.

$$R_i = G(\varepsilon, \sigma) * S_i \quad \text{eq. 3}$$

where i is the index to ring number and varies between 1 and 6.

We used this equation to predict the response to our six rings for a large set of initial pRF parameters; for each vertex, we measured the correlation (our goodness-of-fit index) between the predicted response and the observed t-values. If the highest correlation was $< .7$ the vertex was discarded; otherwise, the parameters yielding the highest correlation were used to initialize a nonlinear search procedure (MATLAB simplex algorithm), which manipulated ε and σ to maximize goodness-of-fit, with the constraint that ε could not exceed 20 deg or be smaller than 1 deg, and σ could not be smaller than .1 deg. Successful fits were obtained for $72.00 \pm 1.86\%$ of V1 vertices, for which the initial coarse grid search gave a correlation > 0.7 and the nonlinear search settled within the constraints. All analyses (on average and distribution of responses and tuning parameters) considered the sub-region of V1 for which a successful fit was obtained. We used ε to estimate the preferred eccentricity of each vertex.

The main modifications of our procedure relative to that described by Dumoulin and Wandell (Dumoulin & Wandell, 2008) are the following: (a) fMRI data were acquired in a block design with only six stimulus types (six eccentricity bands) rather than varying stimulus position at each TR; this allowed us to use a standard GLM approach to estimate each vertex response to the six stimuli (assuming a standard hemodynamic response function) and then use the pRF model to predict these six time-points – much faster than predicting the full fMRI series of 120x2 TRs; (b) our stimuli and consequently our pRFs were defined in one dimension (eccentricity) – whereas the standard pRF is defined in 2D, eccentricity and polar angle (or Cartesian x and y); (c) we maximized the correlation between the predicted and observed fMRI response time-courses rather than minimizing the root mean square error; this eliminates the need to estimate a scale factor to account for the unknown units of the BOLD signal.

Population Tuning for Spatial Frequency

Using a similar logic, we also estimated the population tuning for Spatial Frequency, which represents the aggregate Spatial Frequency tuning of the population of neurons imaged within each vertex area. We modeled the population tuning using a family of functions that includes the

psychophysical Contrast Sensitivity Function (CSF) and can be specified by the following one-parameter equation (Difference-of-Gaussians):

$$pSFT = e^{\frac{-v^2}{\sigma}} - e^{\frac{-v^2}{\sigma/50}} \times \sigma \quad \text{eq. 4}$$

Like we did for the pRF mapping, we defined a stimulus matrix S representing the Fourier spectra of our five bandpass noise stimuli, i.e. the energy of visual stimulation in the frequency domain (here, between 0.03 cpd and 12.5 cpd) for each stimulus. We used the results of our GLM analysis to estimate the vertex response to each of our five bandpass noise stimuli (as t-values; using beta values yields very similar results). We assumed that each vertex response is the linear sum over frequency of the overlap between the pSFT of the voxel and the input stimulus, which is mathematically equivalent to the matrix multiplication between the stimulus and the pSFT.

Like for pRFs, we estimated the best-fit σ parameter of each vertex pSFT with a two-step procedure: a coarse-grid search followed by the simplex search. We used the matrix multiplication of the pSFT and the stimulus to predict the response to our five bandpass noise stimuli for a large set of initial σ values (between 1 and 1,000 in 100 logarithmic steps); for each vertex, we measured the correlation (our goodness-of-fit index) between the predicted response and the observed t-values. If the highest correlation was $< .5$, the voxel was discarded, otherwise the parameter yielding the highest correlation were used to initialize a nonlinear search procedure (MATLAB simplex algorithm), which manipulated σ to maximize goodness-of-fit, with the constraint that σ could not be smaller than .3 and larger than 10,000. Successful fits were obtained for $88.84 \pm 1.28\%$ of V1 vertices for which we obtained a successful eccentricity fit ($86.77 \pm 1.25\%$ of all V1 vertices).

We express the σ parameter in terms of the high-spatial frequency cutoff of the filter (highest spatial frequency at half maximum), $SFCO$ for each vertex:

$$SFCO = 1.26 \sqrt{\frac{\sigma}{2}} - 0.045 \quad \text{eq. 5}$$

Indices defining the effect of deprivation

We computed the effects of short-term monocular deprivation on both the dynamics of binocular rivalry and our fMRI results, estimating the degree to which the two measures are correlated. In all cases, the same equation was applied to psychophysical and fMRI data.

672 The first index, called “Deprivation Index” or DI_{psycho} and DI_{BOLD} is given by eq. 6

673
$$DI = \left(\frac{y_{\text{DepPOST}}}{y_{\text{DepPRE}}} \right) / \left(\frac{y_{\text{NdepPOST}}}{y_{\text{NdepPRE}}} \right)$$
 eq. 6

674 For psychophysics, y = mean duration of Binocular Rivalry phases of the Dep or Ndep eye, during
675 the PRE- or POST deprivation sessions; for fMRI, y = mean BOLD response across V1 vertices to
676 stimuli in the Dep or Ndep eye, during the PRE- or POST-deprivation sessions.

677 The second index, called “Deprived-eye change” or $DepC_{\text{psycho}}$ and $DepC_{\text{cutoff}}$ is given by eq. 7

678
$$DepC = \left(\frac{y_{\text{DepPOST}}}{y_{\text{DepPRE}}} \right)$$
 eq. 7

679 For psychophysics, y = mean duration of Binocular Rivalry phases of the Dep eye, during the PRE-
680 or POST deprivation sessions. For fMRI, y = mean spatial frequency cut-off across V1 vertices
681 estimated for stimuli in the Dep eye, during the PRE- or POST-deprivation sessions.

682
683 *Statistics*

684 Data from individual participants (mean binocular rivalry phase durations or mean BOLD
685 responses/pRF/pST across V1 or V2 vertices) were analyzed with a repeated measure ANOVA
686 approach, after checking that distributions do not systematically deviate from normality by means
687 of the Jarque-Bera test for composite normality (Matlab *jbtest* function, p-values given in the
688 relevant figures). F statistics are reported with associated degrees of freedom and p-values in the
689 Results section, in the form: $F(df, df_{\text{err}}) = \text{value}$; $p = \text{value}$. Post-hoc paired t-tests comparing
690 conditions follow the ANOVA results, in the form: $t(df) = \text{value}$, $p = \text{value}$. Associations between
691 variables are assessed with Pearson product-moment correlation coefficient, reported in the form:
692 $r(n) = \text{value}$, $p = \text{value}$. Aggregate subject data (i.e. vertices pooled across participants and
693 hemispheres) were typically non-normally distributed and thereby were analysed with non-
694 parametric tests. The Wilcoxon sign-rank test was used for comparing medians, and results are
695 reported in the form: $z = \text{value}$, $p = \text{value}$.

696

697 *DATA AND SOFTWARE AVAILABILITY*

698 Data and software are available on Dryad (linked to the current eLife submission).

699

700

701 Figure Legends

702 *Figure 1: Monocular deprivation modulates 7T BOLD responses in early visual cortex*

703 *A: Schematic illustration of the methods. The icons show a band-pass noise stimulus shown to*
704 *either eye through the MR compatible goggles. Before and after the Pre- and Post-deprivation*
705 *scans, outside the bore, we also measured binocular rivalry.*

706 *B: BOLD responses evoked by our band-pass noise stimulus with peak frequency 2.7 cycles per*
707 *degree (cpd), presented in the deprived eye PRE-deprivation, mapped on the flattened cortical*
708 *surface, cut at the calcarine sulcus. T-values are obtained by aligning GLM betas for each subject*
709 *and hemisphere to a left/right symmetric template hemisphere, excluding vertices for which*
710 *preferred eccentricity was not adequately estimated or smaller than 1 (the same criterion used for*
711 *all analyses), then evaluating the distribution of betas in each vertex against 0 (one-sample t-test)*
712 *and FDR correcting across the entire cortical surface. Black dashed lines show the approximate*
713 *average location of the regions of interest V1 through MT, which were mapped on the individual*
714 *subject spaces (see methods); white and blue lines represent the outer limits of the representation of*
715 *our screen space (24 x 32deg) and the foveal representation (≤ 1 deg, where eccentricity could not*
716 *be mapped accurately) respectively.*

717 *C: BOLD modulation during the 3 TRs of stimulus presentation (from 0 to 9s) and the following 4*
718 *blank TRs, for the 2.7 cpd noise stimuli delivered to the deprived eye before deprivation. The y-axis*
719 *show the median percent BOLD signal change in V1 vertices relative to the signal at stimulus onset,*
720 *averaged across subjects. Error bars give s.e. across participants. Note the small between-subject*
721 *variability of the response (given that the response of each subject was computed for just two blocks*
722 *of stimulation-blank).*

723 *D: Average BOLD response to the band-pass noise stimulus with peak frequency 2.7 cpd, in each of*
724 *the four conditions, computed by taking the median BOLD response across all V1 vertices then*
725 *averaging these values across participants (after checking that distributions do not deviate from*
726 *normality, Jarque-Bera hypothesis test of composite normality, all $p > 0.06$). The top black star*
727 *indicates the significance of the ANOVA interaction between factors time (PRE, POST deprivation)*
728 *and eye (deprived, non-deprived); the other stars report the results of post-hoc t-tests: red and*
729 *green stars give the significance of the difference POST minus PRE, for the deprived and non-*
730 *deprived eye respectively; bottom black stars give the significance of the difference deprived minus*
731 *non-deprived eye before and after deprivation. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$; ns non-*
732 *significant.*

733 *E: Histograms of Ocular Drive Index: the difference between the response (GLM beta) to the*
734 *deprived and non-deprived eye, computed for each vertex, separately before and after deprivation.*
735 *Yellow and black lines give the median of the distributions, which are non-normal (logistic) due to*
736 *excess kurtosis.*

737 *Figure 2: Monocular deprivation shifts 7T BOLD Ocular Dominance in V1*

738 *A & C: Average BOLD responses with the same conventions as in Fig. 1D but analysing data from*
739 *two sub-regions of V1. A: only vertices that, before deprivation, respond preferentially to the*
740 *deprived eye. C: only vertices that, before deprivation, respond preferentially to the non-deprived*
741 *eye.*

742 *B & D: Histograms of Ocular Dominance Index (as for Fig. 1E), in the two sub-regions of V1,*
743 *computed before and after deprivation. The black curve simulates the result of adding random noise*
744 *to the distribution obtained before deprivation; only in B does this approximate the distribution*
745 *observed after deprivation.*

746 *Figure 3: Deprivation effects on BOLD and on psychophysics are correlated*

747 *A: Effect of deprivation on Binocular Rivalry dynamics. Average phase duration for the deprived*
748 *and non-deprived eye, before and after deprivation, same conventions as in Fig.1D. Mean phase*
749 *duration distributions do not deviate from normality (Jarque-Bera hypothesis test of composite*
750 *normality, all $p > 0.171$)*

751 *B: Correlation between the deprivation index (the POST to PRE- ratio for the deprived eye divided*
752 *by the same ratio for the non-deprived eye, Eq. 6 in Methods) computed for the binocular rivalry*
753 *mean phase duration and for the BOLD response to our band-pass noise stimulus with peak*
754 *frequency 2.7 cpd. Text insets show the Pearson's correlation coefficient and associated p-value.*

755

756

757 *Figure 4: Deprivation affects spatial frequency selectivity in V1*

758 *V1 BOLD responses to all five of our band-pass noise stimuli (with peaks at 0.1, 0.2, 0.4, 1.1 and*
759 *2.7 cpd, see spectra in Figure 4 - figure supplement 1); A: response to stimuli in either eye, before*
760 *deprivation; B: response to stimuli in either eye, after deprivation. Responses are computed as*
761 *medians across all V1 vertices (like in Fig. 1D), averaged across subjects (error bars report s.e.m.).*
762 *Continuous lines show the response of the best-fit population Spatial Frequency tuning (with the*

one parameter, the high spatial frequency cut-off, indicated in the legend), estimated by applying to the average V1 BOLD response the same model used to predict individual vertex responses (fitting procedure illustrated in Figure 5 - figure supplement 1).

Figure 5: population Spatial Frequency Tuning in V1

A: Maps of pRF eccentricity and best fit spatial frequency cut off (for the deprived eye before deprivation) after aligning the parameter estimates for all hemispheres to a common template and averaging them across subjects and hemispheres, after excluding vertices for which the average preferred eccentricity was not adequately estimated or smaller than 1 (the same exclusion criteria used for analyses).

B: Predicted and observed BOLD activity in one example vertex, elicited in response to our bandpass noise stimuli in the deprived eye PRE (pink) and POST deprivation (red), with best fit spatial frequency cut off (reported in the legend).

C-D: Best fit spatial frequency cut-off, averaged in sub-regions of V1 defined by pRF eccentricity bands, and estimated separately for the two eyes and PRE/POST deprivation.

Figure 6: Deprivation effects on the deprived eye population Spatial Frequency Tuning and binocular rivalry phase duration are correlated.

A: Effect of deprivation on spatial frequency cut off values. Average cut-off across all V1 vertices (pooled across eccentricities) for the deprived and non-deprived eye, before and after deprivation, same conventions as in Fig. 1D. Distributions of the log-values do not deviate from normality (Jarque-Bera hypothesis test of composite normality, all $p > 0.285$).

B: Correlation between the POST/PRE ratio (Eq. 7 in the Methods) computed for the binocular rivalry mean phase duration and for the spatial frequency cut off for the deprived eye.

Figure 7: Deprivation effects are stronger in ventral than in dorsal stream areas.

Panels A-B show V4 responses across spatial frequency stimuli presented to each eye (colored lines) before (A) and after deprivation; panels C-D show V3a responses and panels E-F show hMT+ responses. Each data point is computed by taking the median BOLD response across vertices in the region of interest for each stimulus and subject, then averaging across subjects (errorbar report s.e.m.).

Panel G summarizes the effect of deprivation measured for the highest spatial frequency stimulus in the V1, V2, V3/VP, V4, V3a and hMT+ region of interest, computing the interaction term (POST-

793 *PRE difference of BOLD response for the deprived eye, minus the same value for the non-deprived*
794 *eye) for individual participants and the 2.7 cpd stimulus. Values around 0 indicate no effect of*
795 *deprivation and values larger than 0 indicate a boost of the deprived eye after deprivation. One-*
796 *sample t-tests comparing this value against 0 give a p-value equivalent to that associated with the*
797 *interaction term of the ANOVA (Fig. 1D); the significance of the resulting t-value is given by the*
798 *stars plotted below each errorbar. Stars plotted above the lines show the results of paired t-tests*
799 *comparing interaction terms in V4 and V3a/hMT+. *** = $p < 0.001$; ** = $p < 0.01$; * = $p < 0.05$; ns*
800 *= $p \geq 0.05$. Green and Blue highlight the assignment of the higher tier areas to the ventral and*
801 *dorsal stream respectively.*

802 [Supplementary material](#)

803 [Figure 1 - figure supplement 1: Effects of deprivation across the visual cortex](#)

804

805 *Monocular deprivation had strong and opposite effects on the response to the 2.7 cpd stimulus in*
806 *the two eyes. Panels in the top row report responses to the non-deprived eye, those in the bottom*
807 *row to the deprived eye. The central panels map the average %BOLD response to stimuli presented*
808 *in either eye, before and after monocular deprivation (isoeccentricity lines are taken from the pRF*
809 *mapping shown in Fig. 5A main text), showing that suppression of the non-deprived eye and*
810 *enhancement of the deprived eye are largely homogeneous within each ROI. Panel A & G compare*
811 *% BOLD responses in each of our 19 individual participants (each point shows the average across*
812 *the 2 hemispheres), to stimuli in the non-deprived eye, before vs. after deprivation; the majority of*
813 *points lie below the bisection of the axes, implying a reduction of responses to the non-deprived eye*
814 *after monocular deprivation. The same comparison for stimuli in the deprived eye in panels B & H*
815 *shows an increase of BOLD responses: most point lie above the bisection line, implying a boost of*
816 *responses to the deprived eye after deprivation.*

817 [Figure 1 - figure supplement 2: Change of ocular preference after deprivation](#)

818 *Panels A & B map the difference of the %BOLD response to the Deprived – Non deprived eye,*
819 *before and after deprivation, respectively. While there is no organized preference for either eye in*
820 *any visual area before deprivation, there is a clear preference for the deprived eye after deprivation*
821 *(the net result of the boost of the deprived eye response and the suppression of the non deprived eye*
822 *response), which spreads across most of the areas activated by our stimulation.*

823 [Figure 1 - figure supplement 3: Split-half reliability of the deprivation effect in V1](#)

824

825 *Average V1 BOLD response to the band-pass noise stimulus with peak frequency 2.7 cpd, same as*
826 *in Figure 1D but computed separately for each of the two stimulus repetitions that occurred in each*
827 *scan (PRE and POST, to either eye). This essentially splits the dataset in half, and we show that*
828 *both halves reveal a significant interaction between the factors time (PRE, POST deprivation) and*
829 *eye (deprived, non-deprived; interaction term: first block $F(1,18) = 7.53470$, $p = 0.01332$, second*
830 *block $F(1,18) = 7.11116$, $p = 0.01572$).*

831 *Figure 4 - figure supplement 1: Bandpass noise stimuli*

832 *A: example time-course of stimulation, showing the blocked presentation of the five spatial*
833 *frequency stimuli. Blocks were presented in pseudo-random order, twice per run, for a total of 70*
834 *TRs. In each run, stimuli were only presented to one eye, while the other was shown a mid-level*
835 *gray screen. Each eye was tested once, before and after deprivation.*

836 *B. example of the 2D bandpass noise stimuli, with their effective RMS contrast.*

837 *C: Normalized spectra of the bandpass filter that, multiplied by white noise, generated the five*
838 *bandpass noise stimuli.*

839

840 *Figure 5 - figure supplement 1: population Spatial Frequency Tuning estimation*

841 *A: family of functions used to model spatial frequency sensitivity in individual vertices. Different*
842 *curves are generated by manipulating a single parameter, which is linearly related to the high*
843 *spatial frequency cut-off of the function.*

844 *B: spatial frequency tuning curves in A were multiplied by the five spatial frequency spectra*
845 *defining our band-pass noise stimuli, yielding a five-element vector that predicts the BOLD*
846 *response to the stimuli.*

847 *C-I: The BOLD response observed in each vertex (or pool of vertices, for Fig. 4) was fit with the*
848 *model, by varying the spatial frequency cut-off and finding the value for which the predicted BOLD*
849 *response correlates best with the observed BOLD response. For the example vertex in panel C, the*
850 *best fit cut-off is 0.5 cpd.; panels D-I show individual vertices for which the best fit cut-off is 1, 2, 4,*
851 *8, 16 or 32 cpd (see text insets).*

852

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864 [Competing of Interests](#)

865 The authors declare no competing interests.

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870 [References](#)

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Figure 1: Monocular deprivation modulates 7T BOLD responses in early visual cortex

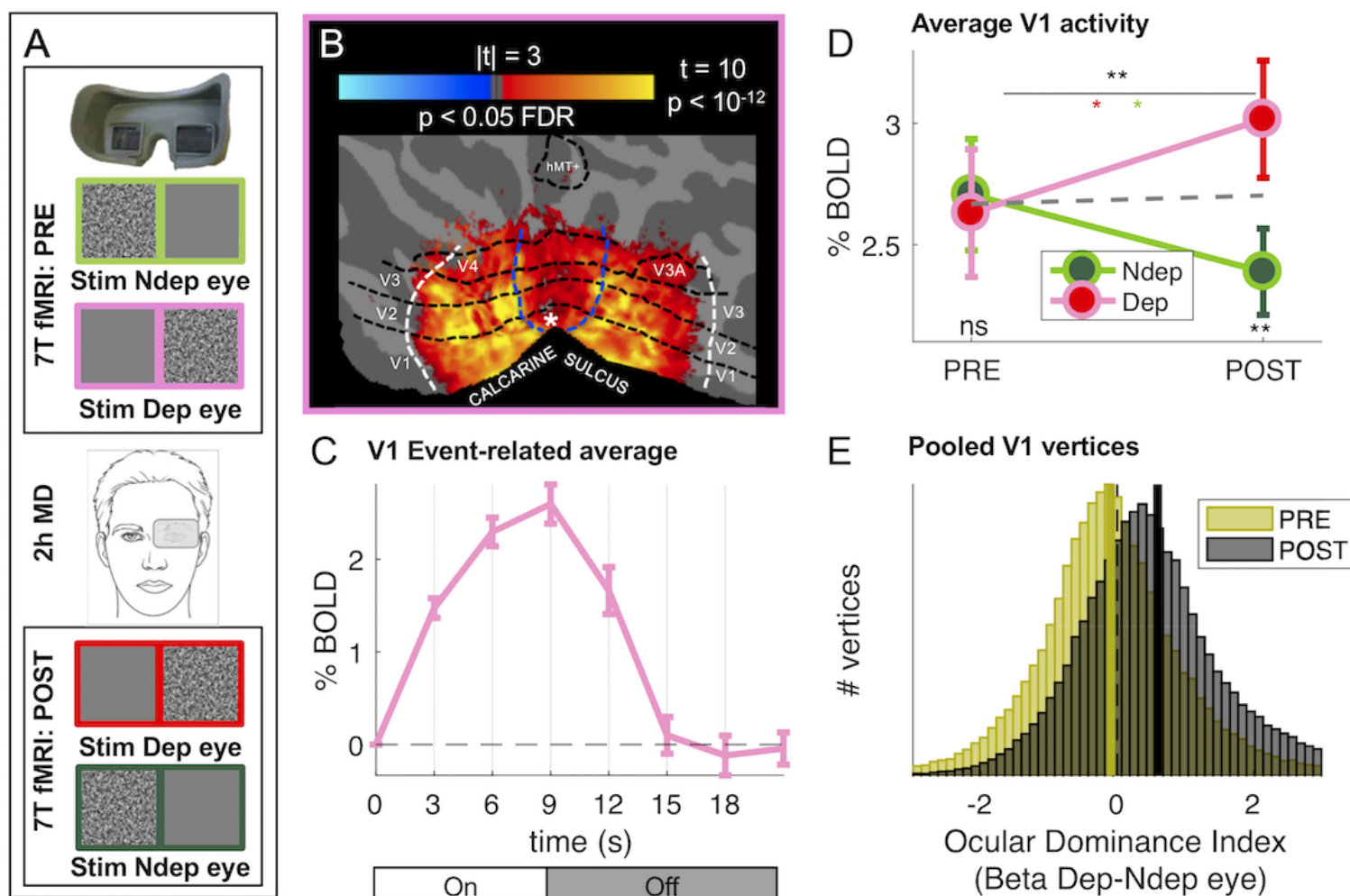


Figure1-figuresupplement1: Effects of deprivation across the visual cortex

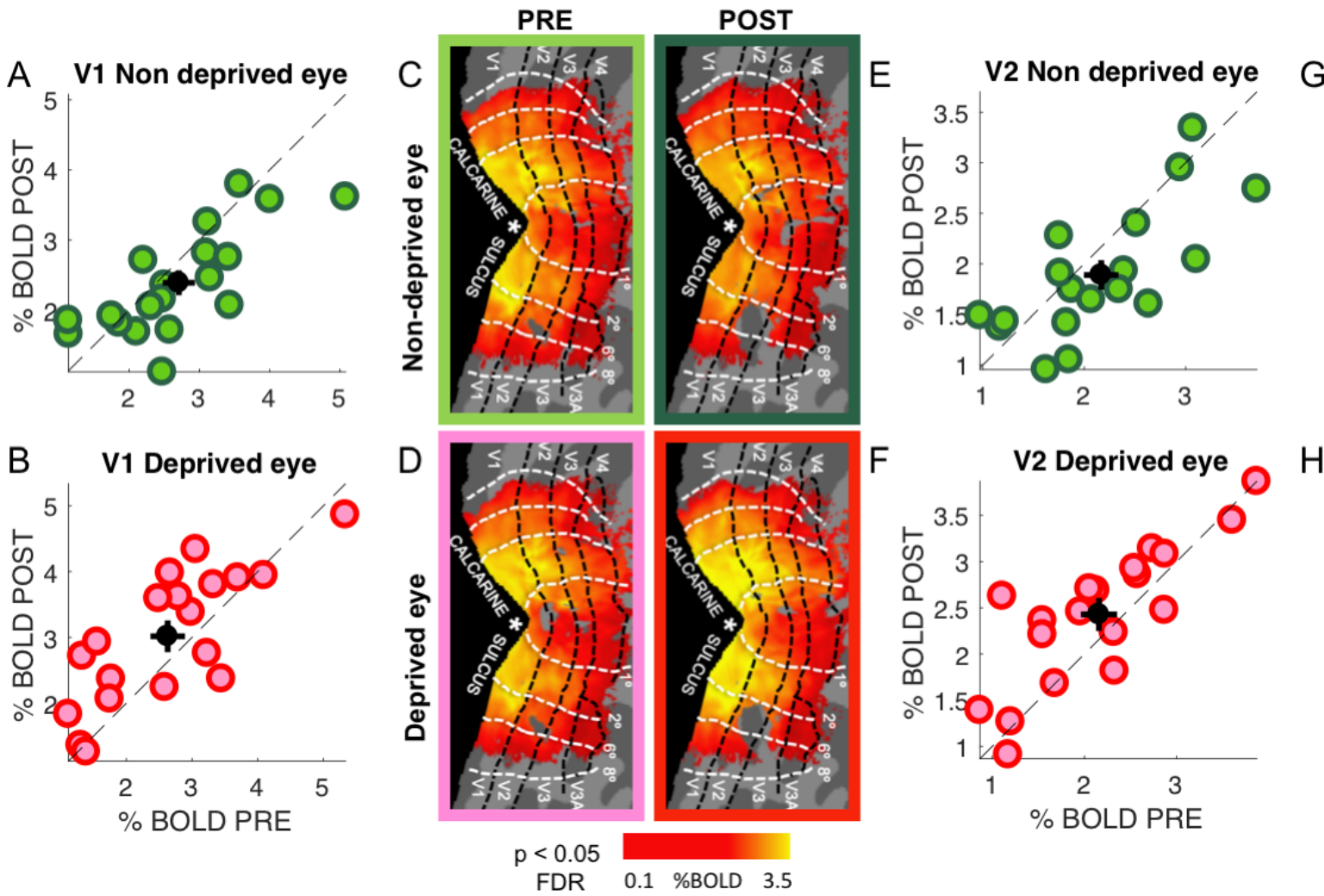


Figure1-figuresupplement2: Change of ocular preference after deprivation

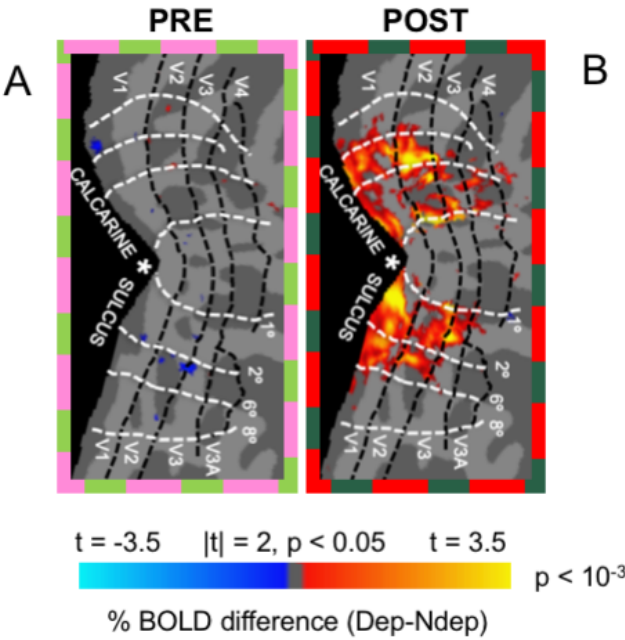


Figure1-figuresupplement3: Split-half reliability of the deprivation effect in V1

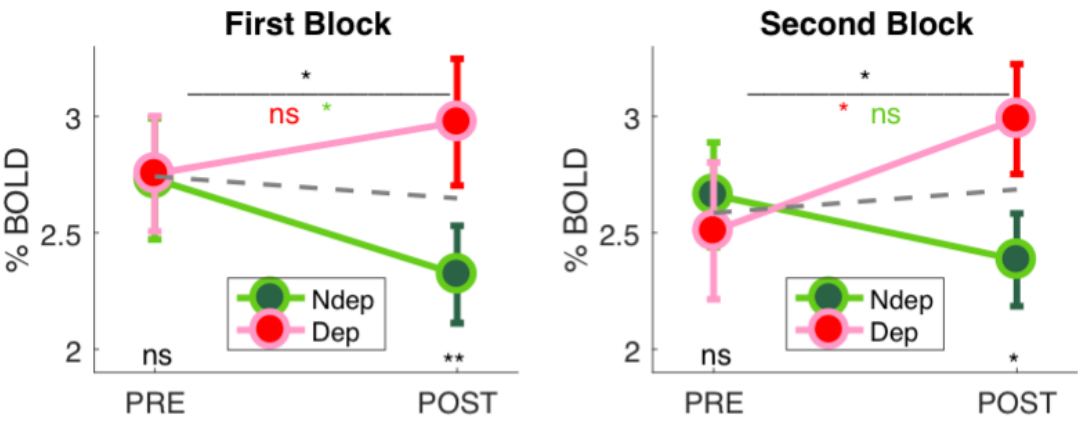
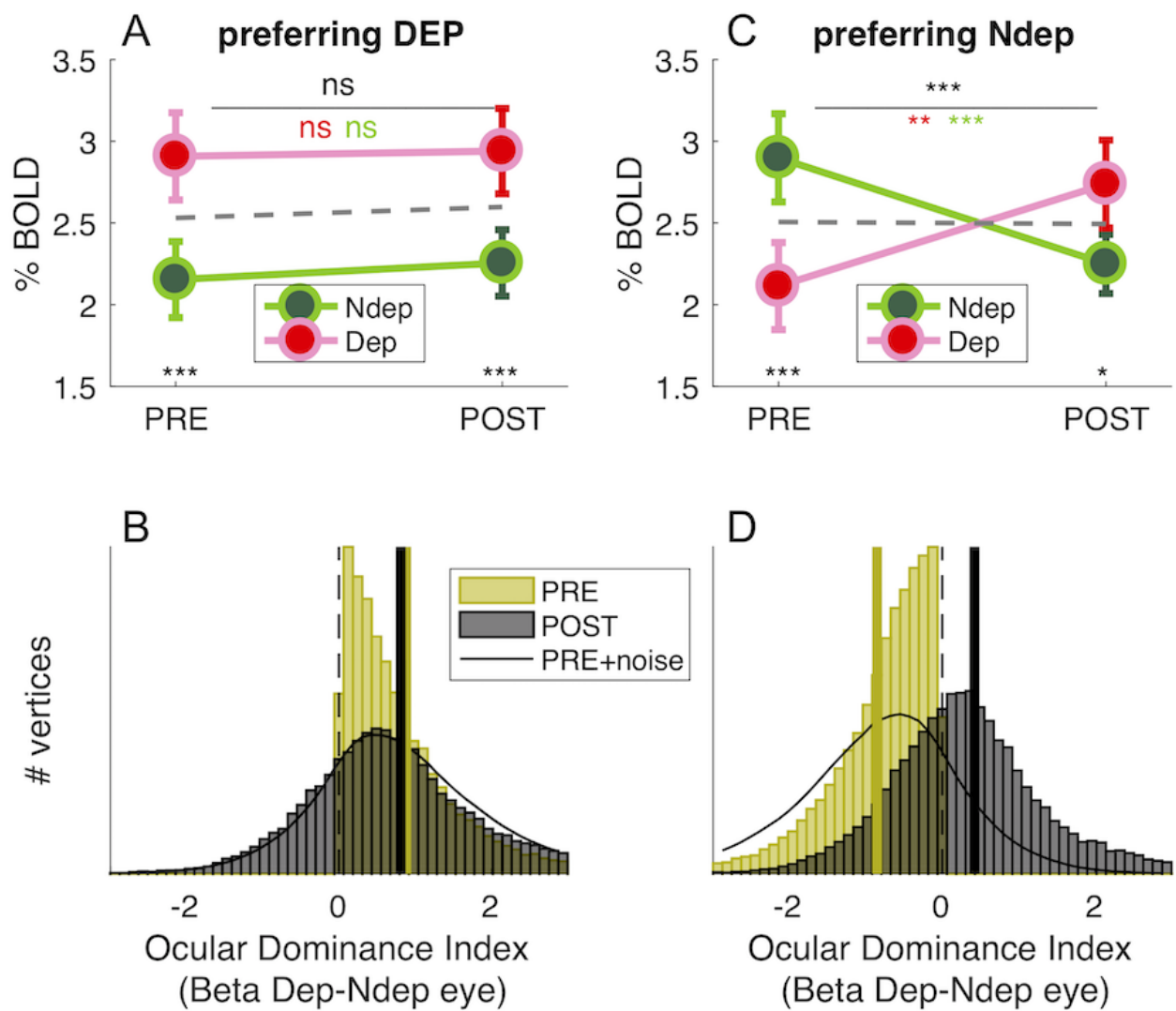


Figure 2: Monocular deprivation shifts 7T BOLD Ocular Dominance in V1



PSYCHOPHYSICS
Binocular Rivalry

A

Mean phase duration (s)

PRE POST

Ndep

Dep

CORRELATION
% BOLD - Binocular Rivalry

B

Deprivation index on % BOLD

Deprivation index Bin Riv

$R(19)=0.48^*$
 $p=0.036$

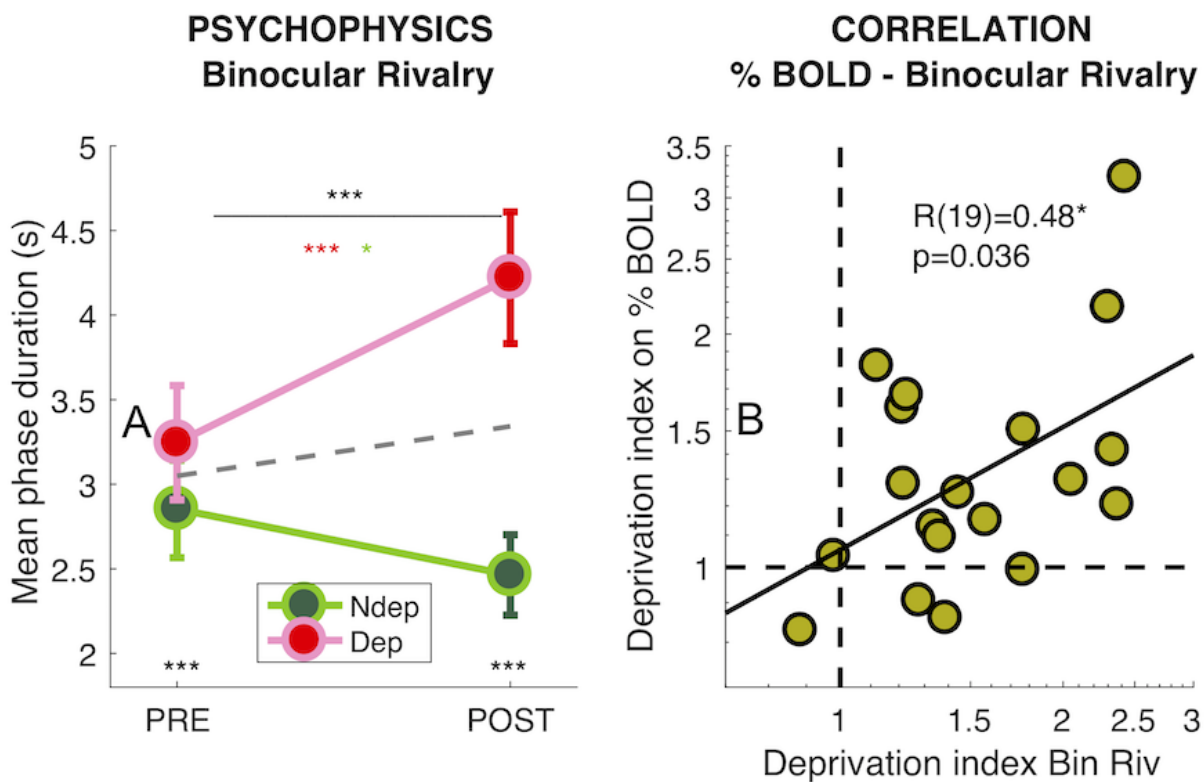


Figure 4: Deprivation affects spatial frequency selectivity in V1

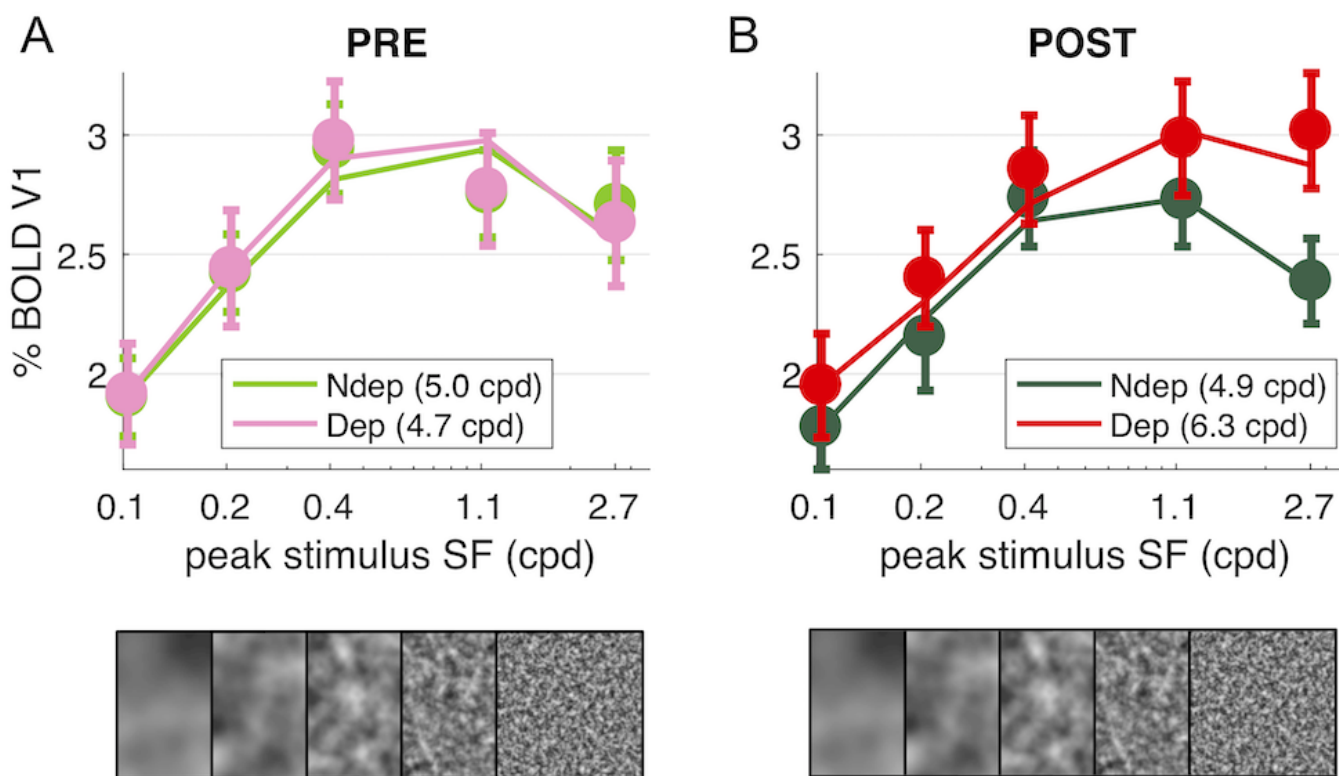


Figure4-figuresupplement1: Bandpass noise stimuli

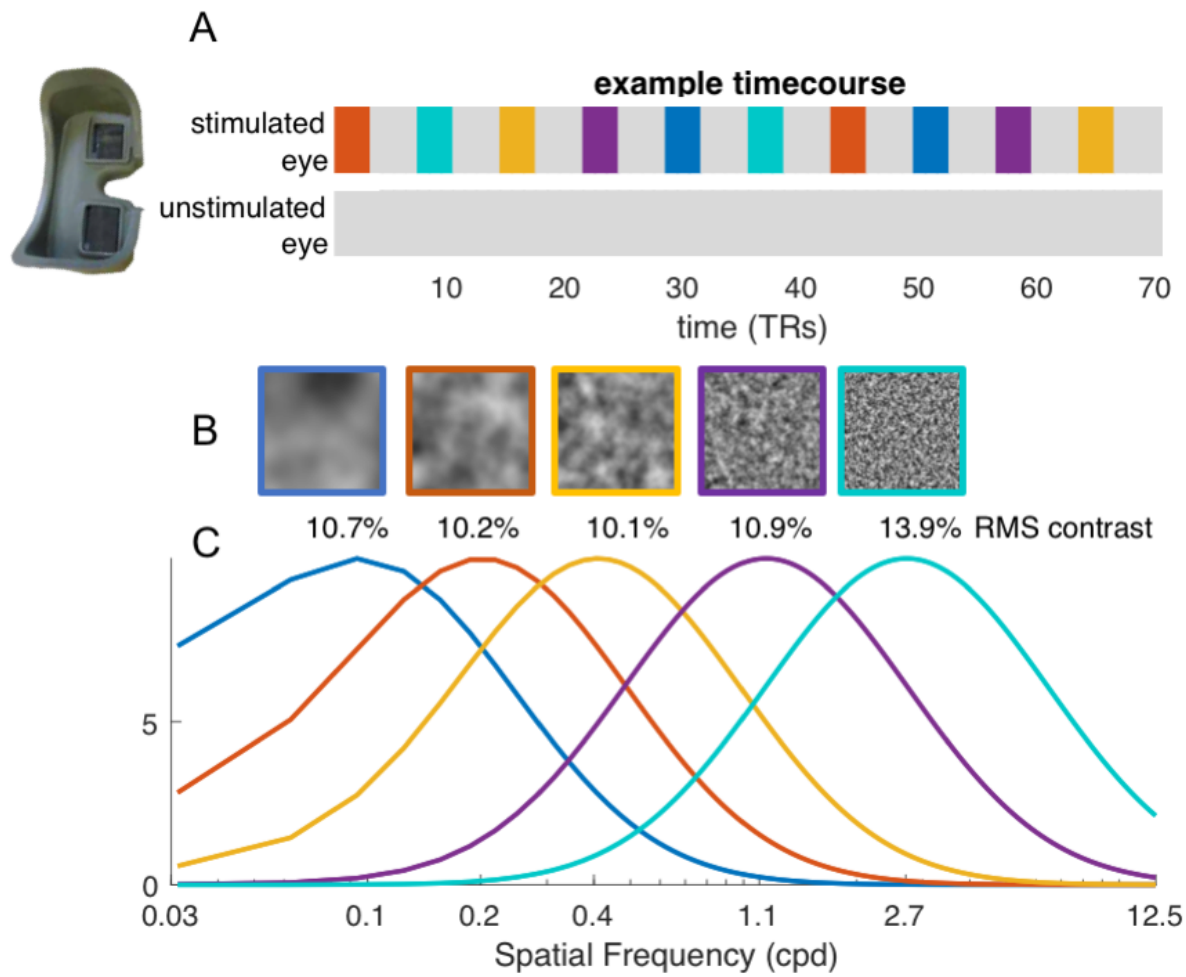


Figure 5: population Spatial Frequency Tuning in V1

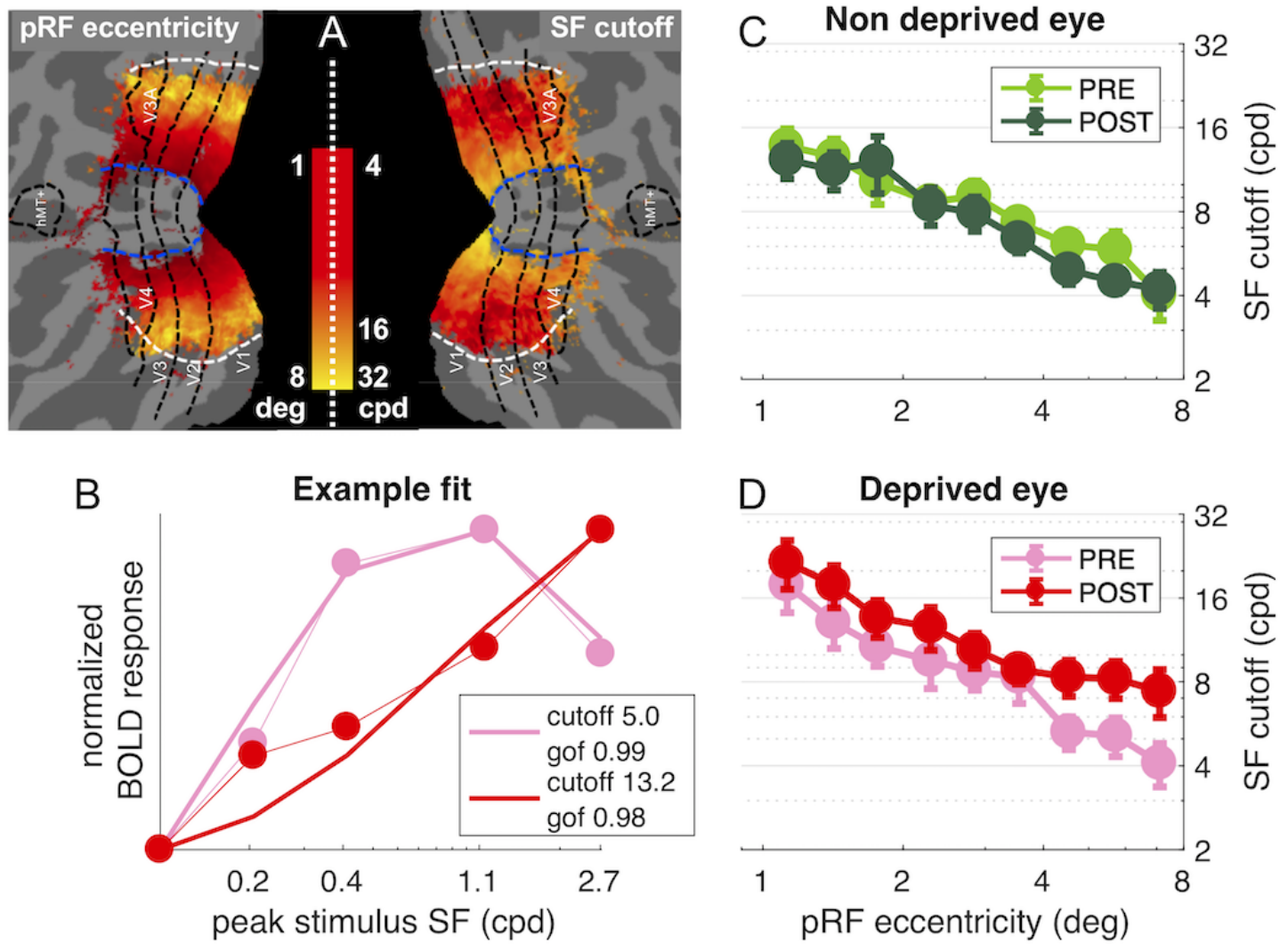


Figure5-figuresupplement1: population Spatial Frequency Tuning estimation

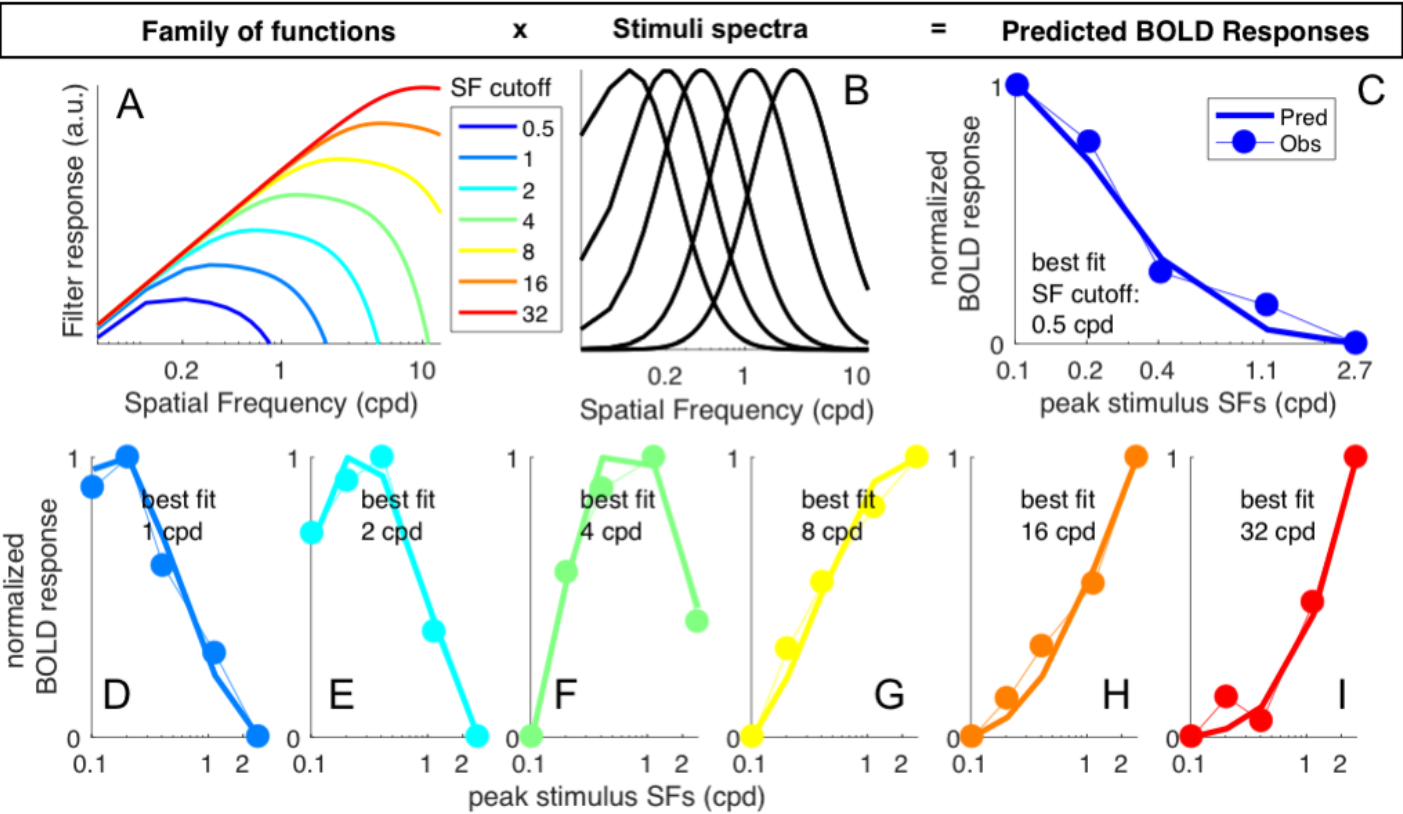


Figure 6: Deprivation effects on the deprived eye population Spatial Frequency Tuning and binocular rivalry phase duration are correlated.

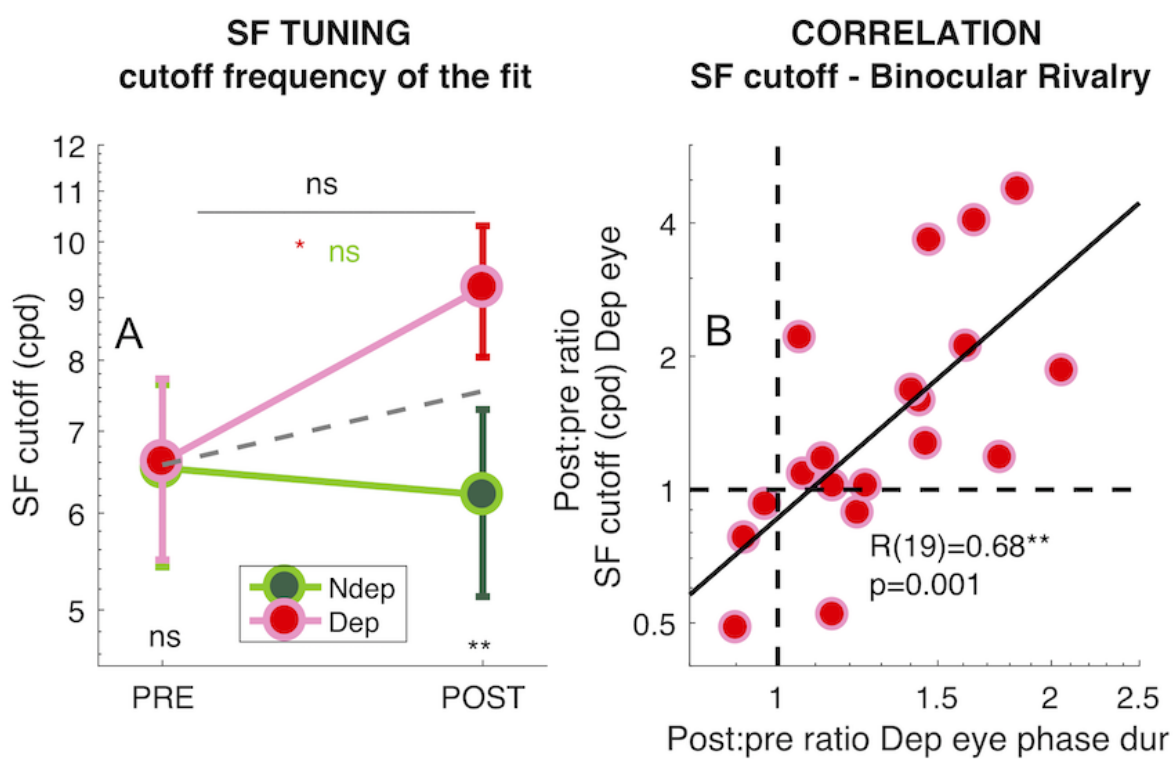


Figure 7: Deprivation effects are stronger in ventral than in dorsal stream areas.

